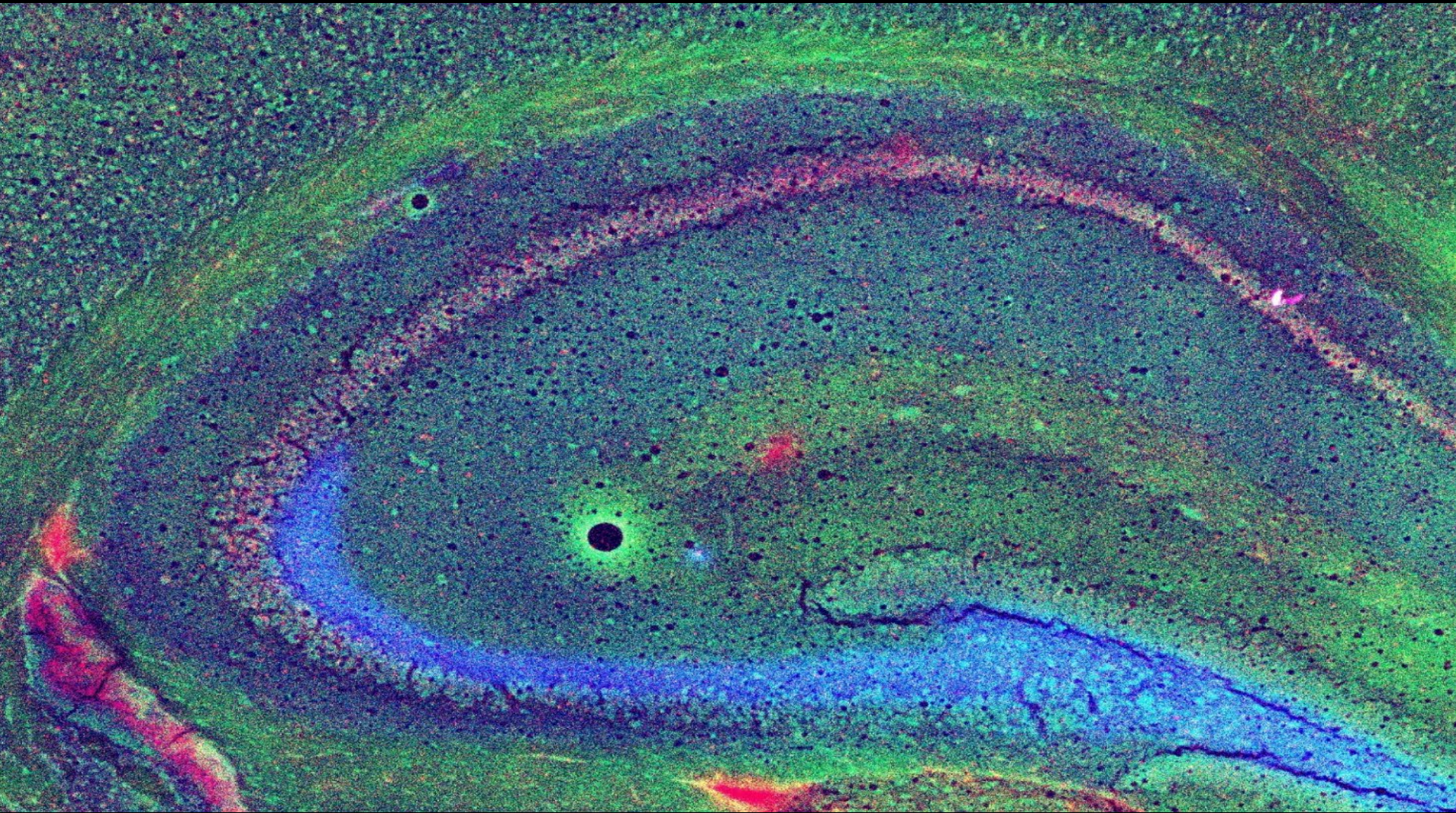


Sample Preparation Considerations for Multi-Modal IRM and XFM Analyses of Biological Tissues



IRM & XFM Workshop, May 20th 2021
Dr Mark Hackett – Curtin University



Curtin University

BACKGROUND

- ❖ BSc (Hons) Analytical Chemistry (Curtin University, 2006)
- ❖ PhD Chemistry (The University of Sydney, 2007 - 2010)
 - *Spectroscopic investigation of cerebral malaria*
 - *ischemia and blood-brain-barrier*
- ❖ Research Fellowships in Canada (University of Saskatchewan 2011 - 2015)
 - *Spectroscopic studies of neurodegeneration after stroke*
 - *ischemic stroke, intra-cerebral haemorrhage*
- ❖ Senior Lecturer in Analytical Chemistry at Curtin University (2016)
 - *Developing and applying spectroscopic approaches to study chemical mechanisms of memory loss during ageing and dementia*



Analytical Chemistry
Applied Spectroscopy

Neuroscience
Neurochemistry



CURTIN HEALTH
INNOVATION
RESEARCH INSTITUTE



Curtin University



Australian Government

Australian Research Council

CHEMISTRY @ CURTIN



TALK OUTLINE



❖ Themes

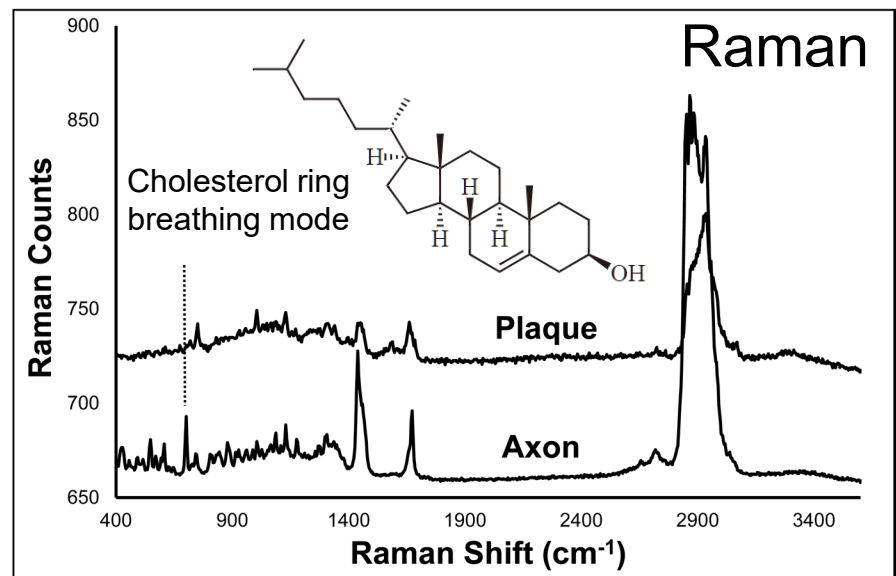
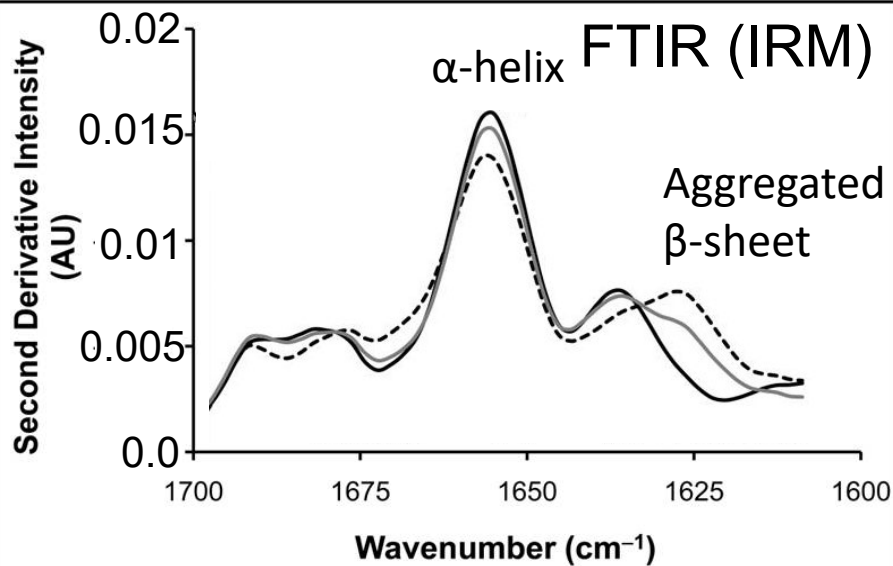
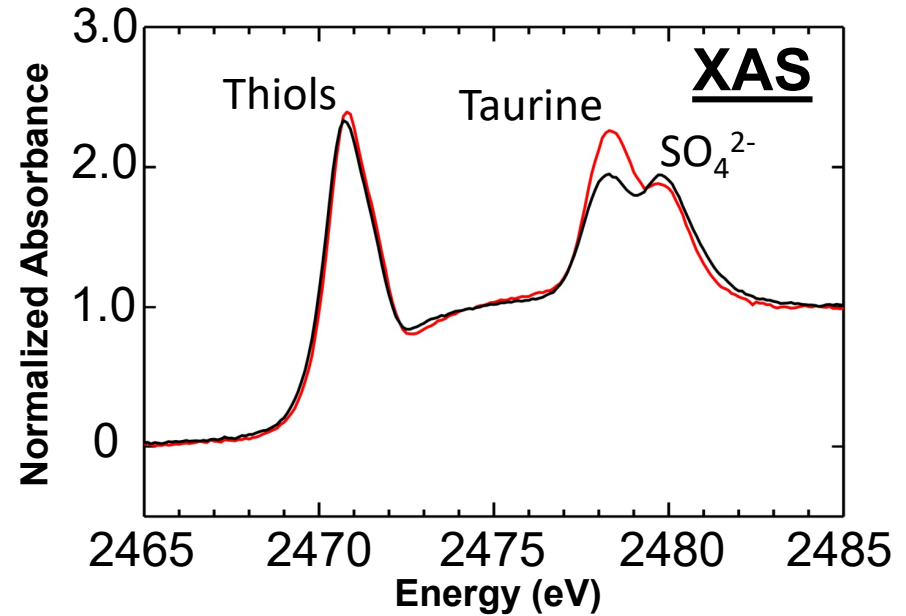
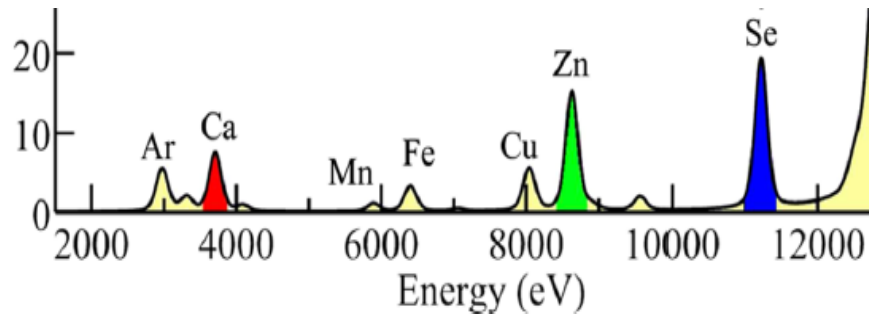
- ❖ Techniques
- ❖ Sample Preparation
- ❖ Combined Histology or Immuno-histochemistry with XFM or IRM

❖ Research Examples

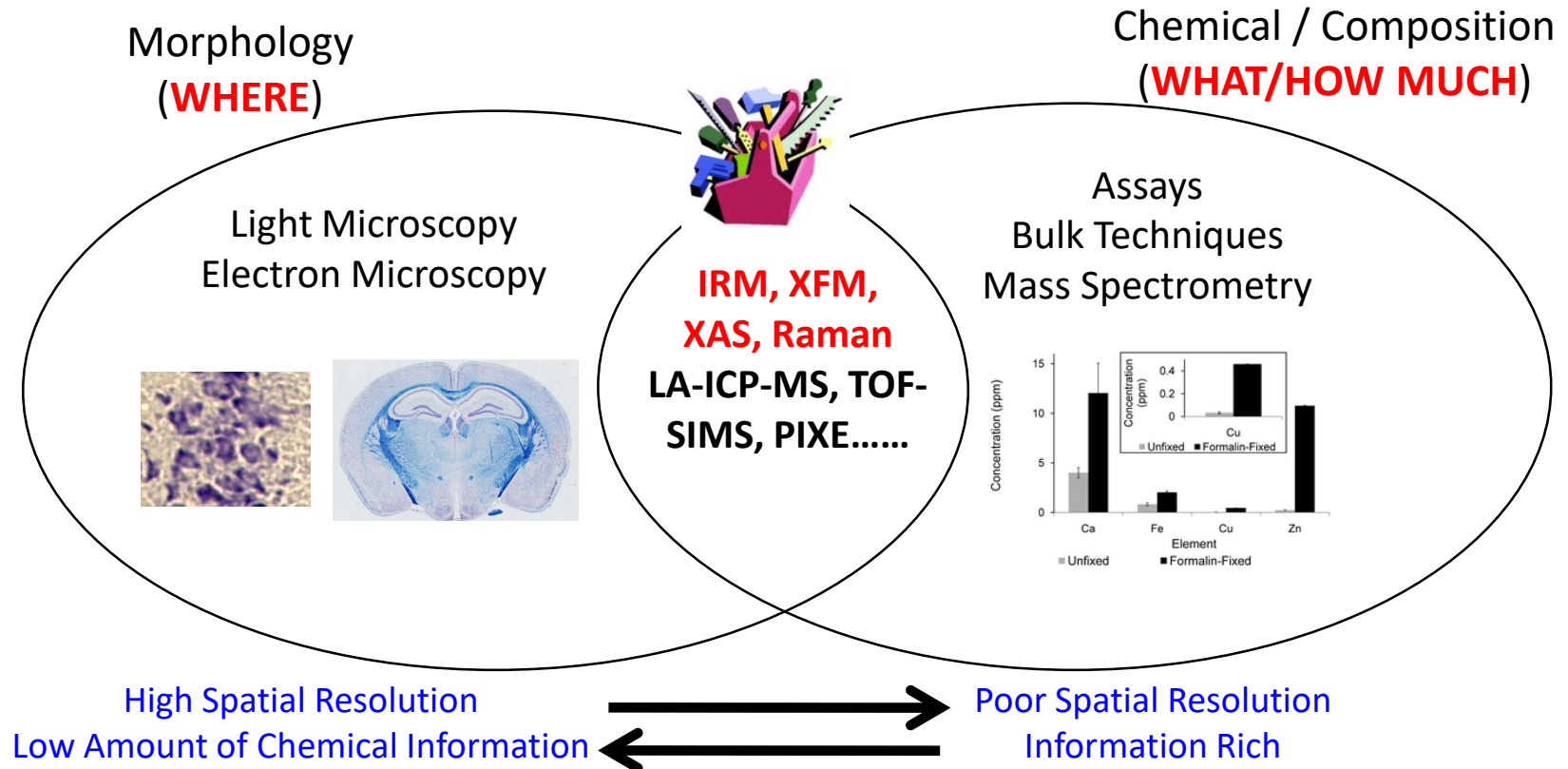
1. Colocalisation of metal ions with macromolecules in amyloid plaques
2. Brain-Fe homeostasis during natural ageing
3. XANES imaging to study hippocampal Zn speciation
4. Medium energy XAS to study sulfur speciation after stroke

INTRODUCTION: THE TECHNIQUES I USE.....

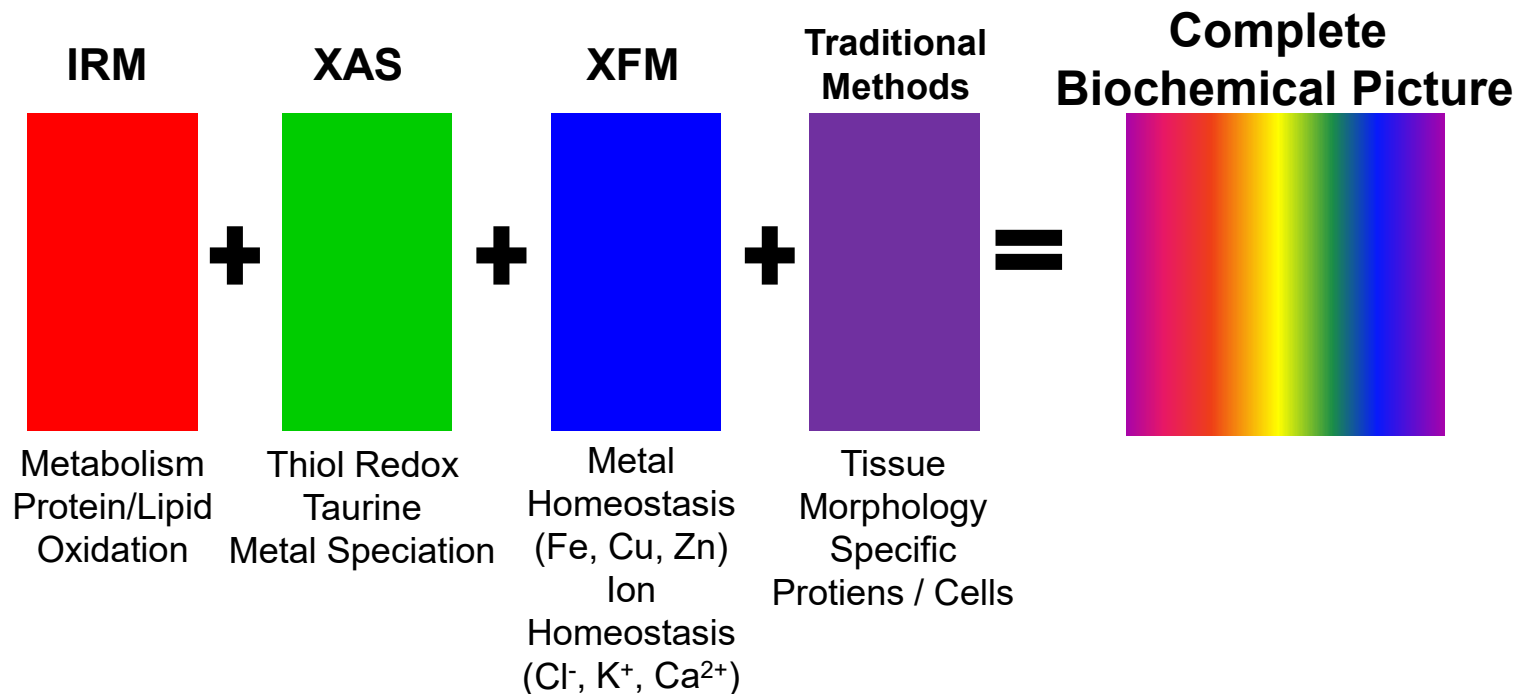
XRF (XFM)



WHAT IS THE ROLE OF SPECTROSCOPIC IMAGING?



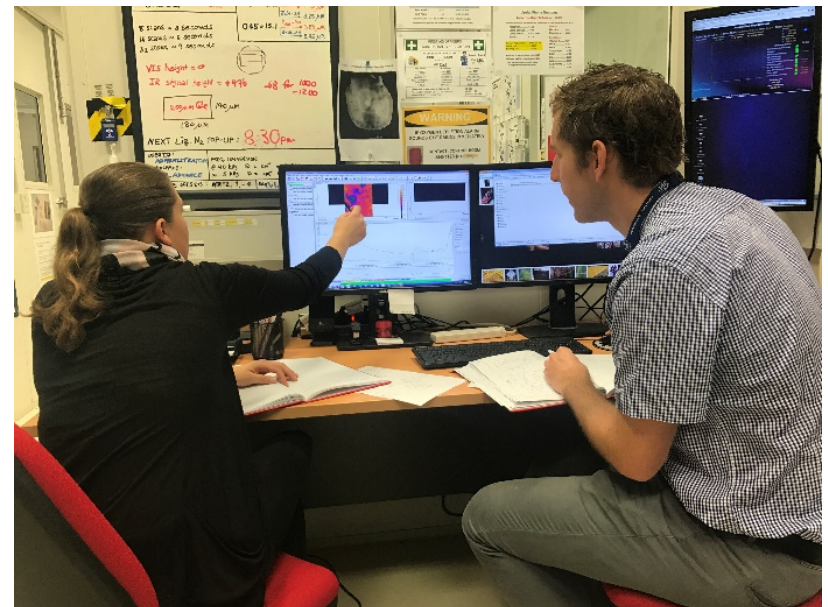
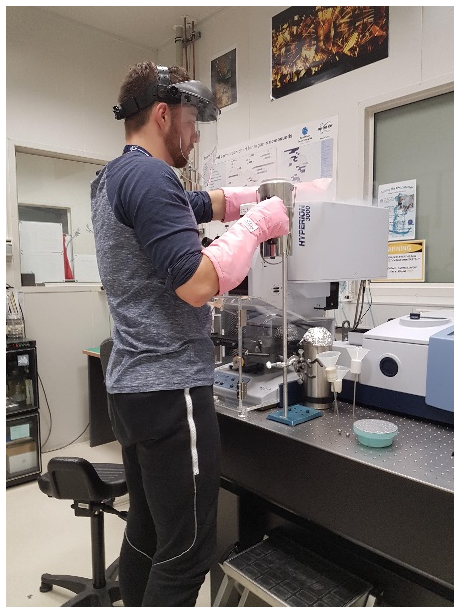
THE MULTI-MODAL APPROACH



WHY THE SYNCHROTRON??

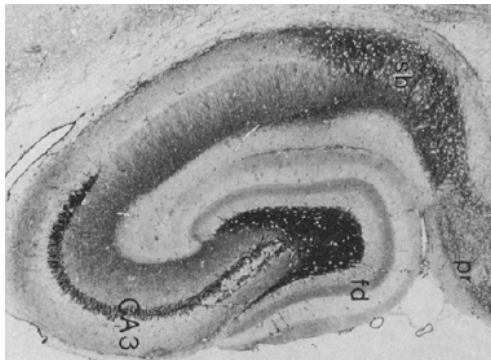


A bright & tuneable light source!

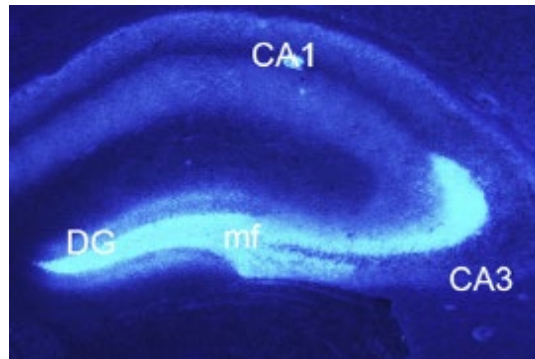


METAL HOMEOSTASIS AND BRAIN FUNCTION

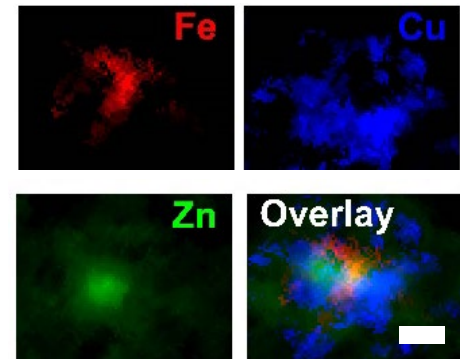
- Zinc binding accelerates amyloid- β plaque formation
- Zn deficiency is associated with memory loss
- Brain Fe accumulates during ageing and neurodegenerative disease
- **How do we study brain-metal homeostasis?**



Danscher et. al.,
Histochemistry 55, 27-40. **1978**



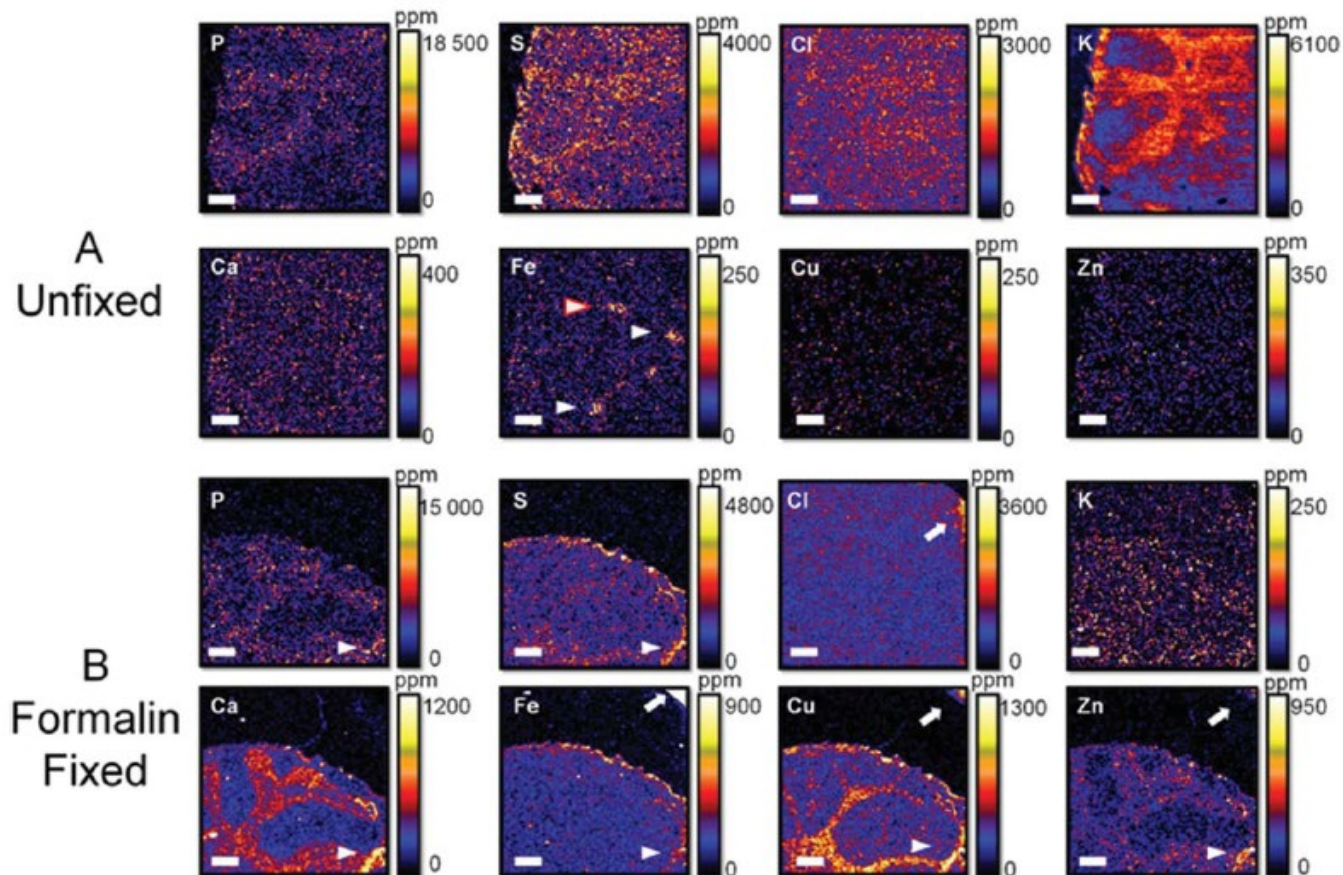
Lee, Joo-Yong, et. al.,
Brain research 1418, 12-22, **2011**



Summers et. al.,
Biochemistry, 56, 4107-4116 **2017**

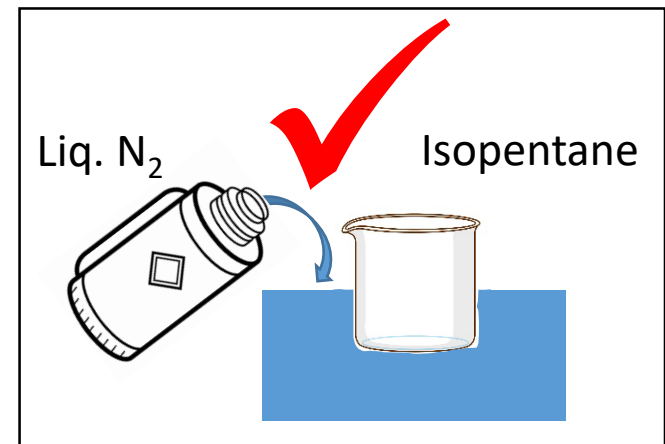
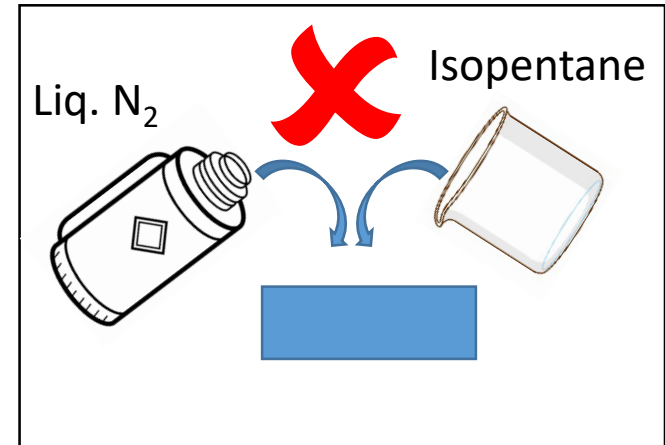
EXPERIMENTAL CONSIDERATIONS FOR XFM/IRM REALLY IMPORTANT!

- Preparing tissues for XFM and IRM requires different preparation than histology and immuno-histochemistry



SAMPLE PREPARATION – RAPID FLASH FREEZING

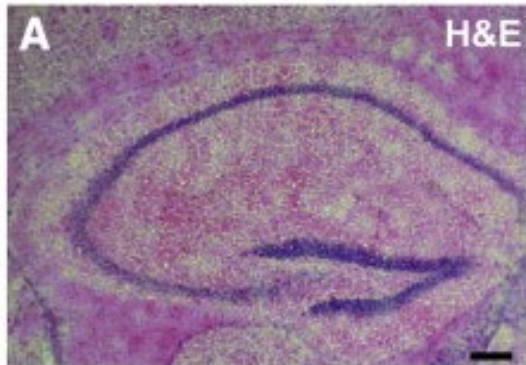
- Tissues samples are rapidly dissected and immediately flash frozen in liquid-nitrogen cooled iso-pentane
 - *(no perfusion, no fixation)*
- Samples stored in -80°C until ~ 1 week before analysis
- Samples sectioned on a *cryo-microtome at -16°C* for brain (liver -18°C), *$10\text{-}\mu\text{m}$ -thick*. This is warmer than most SOPs, which are written for sucrose-cryoprotected, formalin-fixed tissue
- Tissue sections “melted” onto substrate, and either *transferred back to -80°C freezer, or air-dried*
- Analysis typically within 1 week of sectioning
- If your samples are cracking, or tearing when you section, they are too cold. It takes about 30 – 40 min for tissue to warm from -80°C to -16°C in cryostat
- Substrates
 - 500 nm, 1 μm , or 2 μm thick Si_3N_4
 - *XFM, or multi-modal XFM/IRM*
 - 500 μm or 1 mm thick CaF_2 , BaF_2
 - *For IRM*



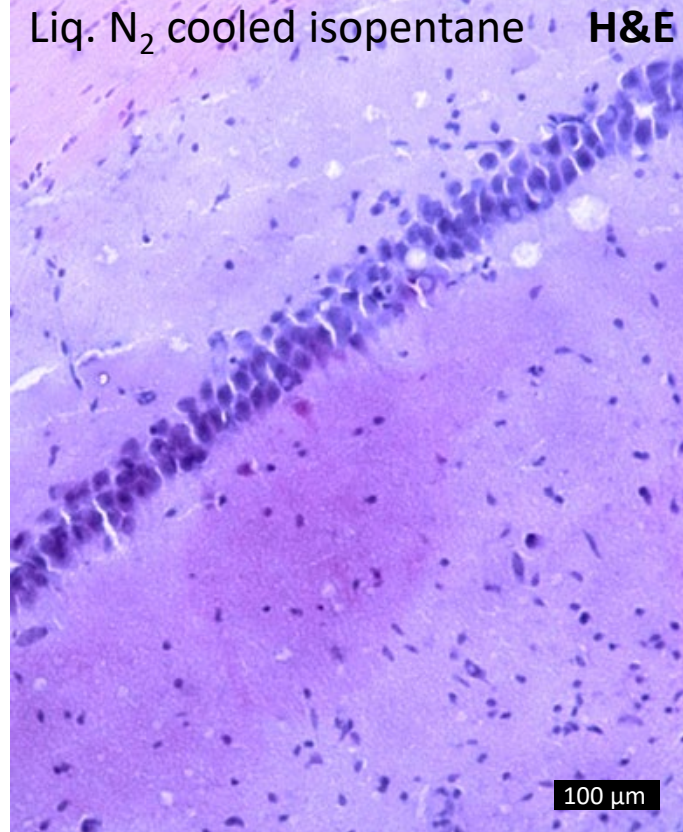
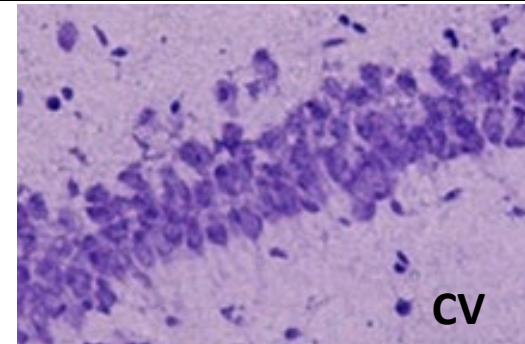
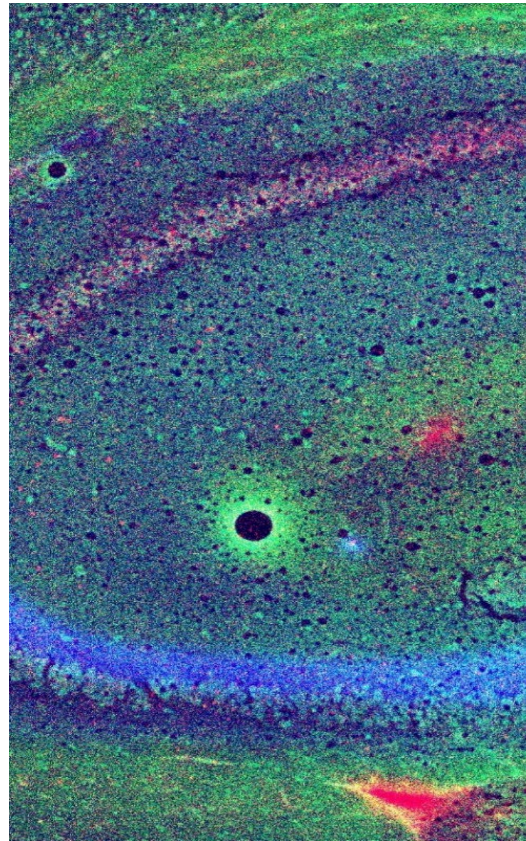
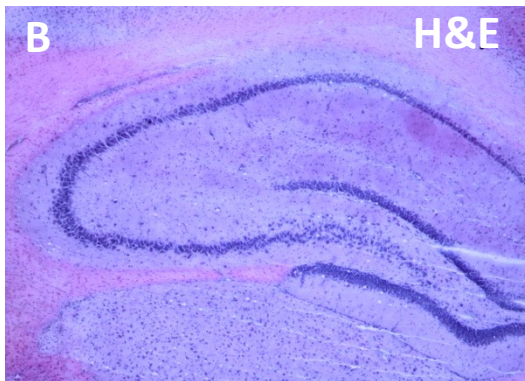
RAPID FLASH FREEZING, HISTOLOGY, & ICE CRYSTALS

- Histology and immuno-histochemistry can easily be performed on flash frozen tissues
- Either on the same tissue analysed at synchrotron, or serial sections
- Post-sectioning immersion fixation (typically 10 % PFA)

Freezing on dry ice

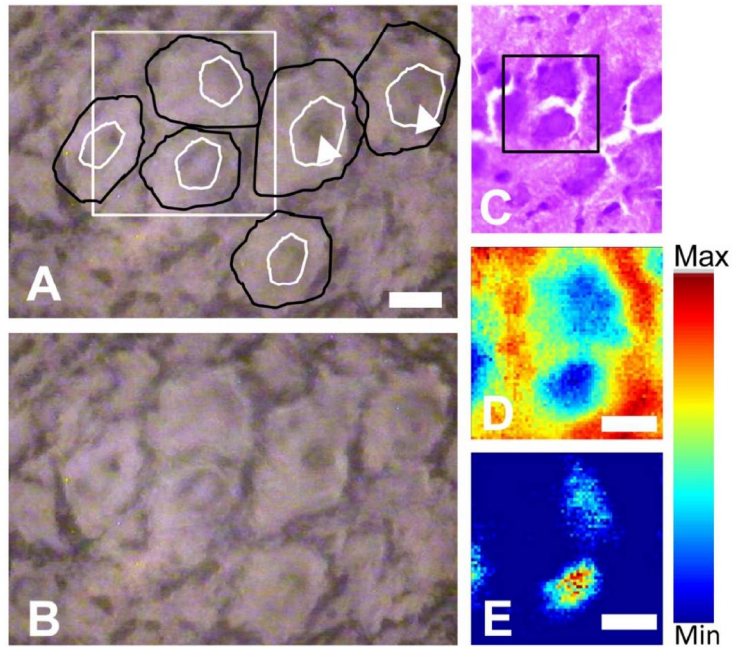


Liq. N₂ cooled isopentane

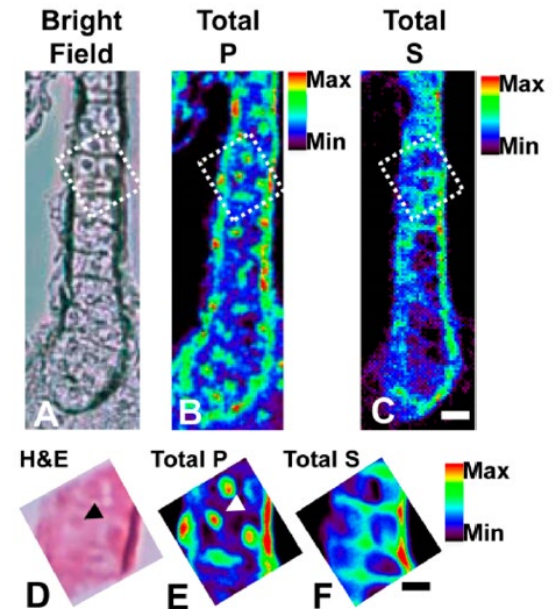
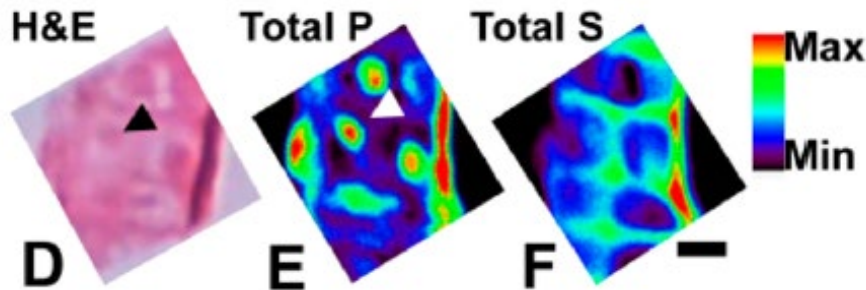
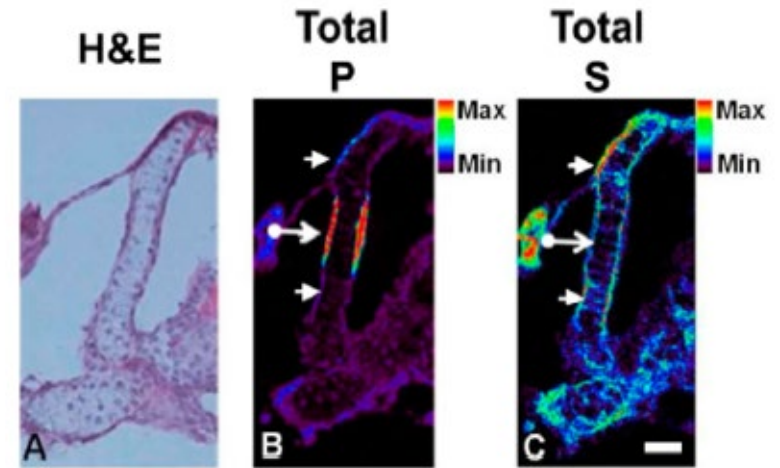


SYNCHROTRON IMAGING, HISTOLOGY (SAME SAMPLE)

Optical Microscopy on Unstained Tissue, IRM, and H&E staining (CaF_2)

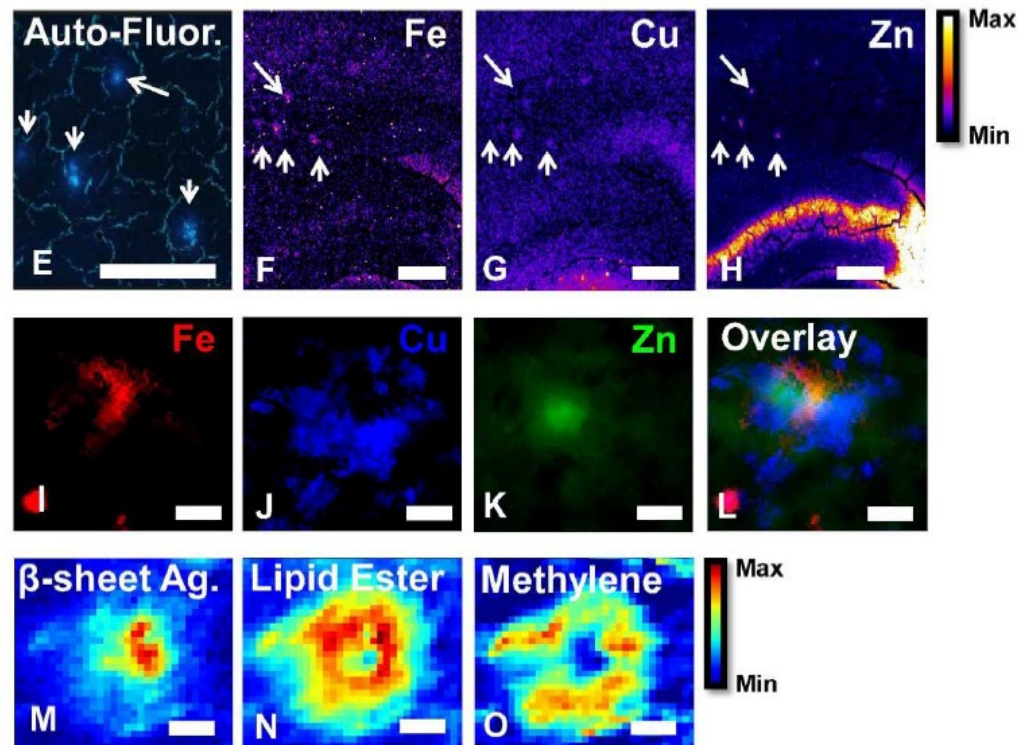


XFM (nano-probe) and H&E Histology (Si_3N_4)



MULTI-MODAL INVESTIGATION OF A β -PLAQUES

- Is there an association between Cu and lipids in plaque periphery?
- UV-autofluorescence, XFM (metal ions), IRM (protein aggregates, lipids)
- 10- μ m-thick, air-dried tissue sections, Si₃N₄



IMAGING BRAIN IRON HOMEOSTASIS DURING AGEING

- Brain Fe increases (if you compare adult to senescent animals)
- Fe is redox active, does elevated brain Fe contribute to heightened oxidative stress?
- **Where is Fe accumulating in the brain, and in which cell types?**

WHY NOT JUST USE PERLS' IRON HISTOCHEMISTRY?

Reaction of Ferric Iron with Ferrocyanide

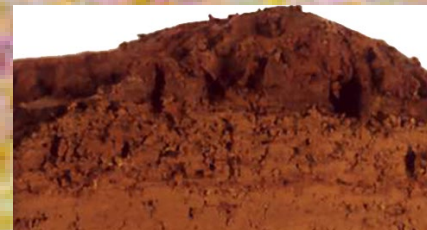
Perls Stain: $4\text{Fe}^{3+} + 3[\text{Fe}(\text{CN})_6]^{4-} \longrightarrow \text{Fe}^{\text{III}}[\text{Fe}^{\text{III}}\text{Fe}^{\text{II}}(\text{CN})_6]_3$

Requires dilute acid

Does not stain all forms of iron (no heme, no labile Fe)

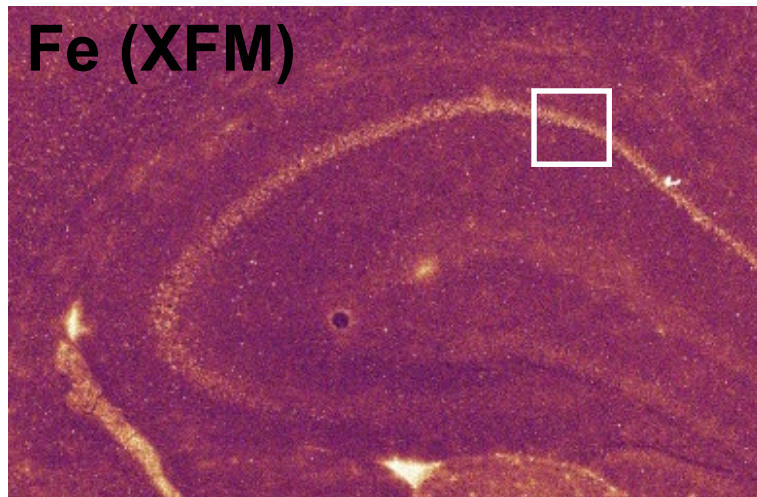
Not quantitative

Ferrihydrite
 $(\text{Fe}^{3+})_2\text{O}_3 \cdot 5\text{H}_2\text{O}$

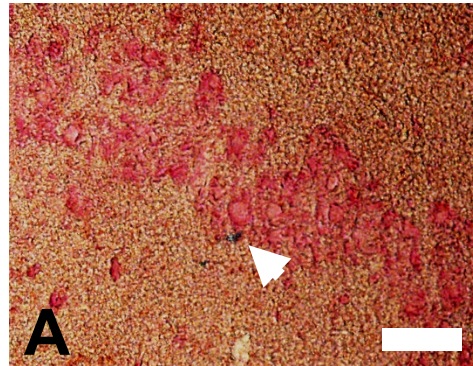


ARE NEURONS ENRICHED IN IRON? (YES)

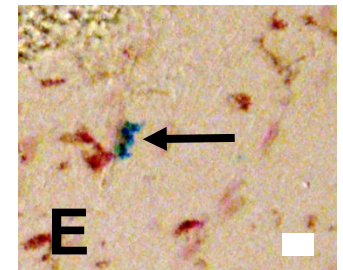
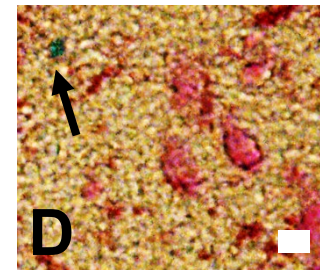
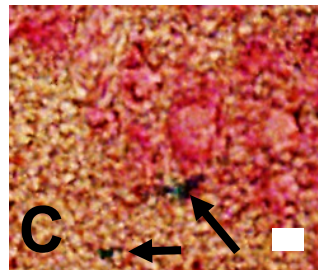
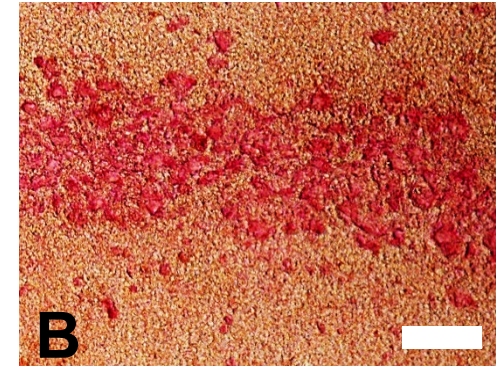
Conflicting results between elemental mapping and Perl's histochemistry?



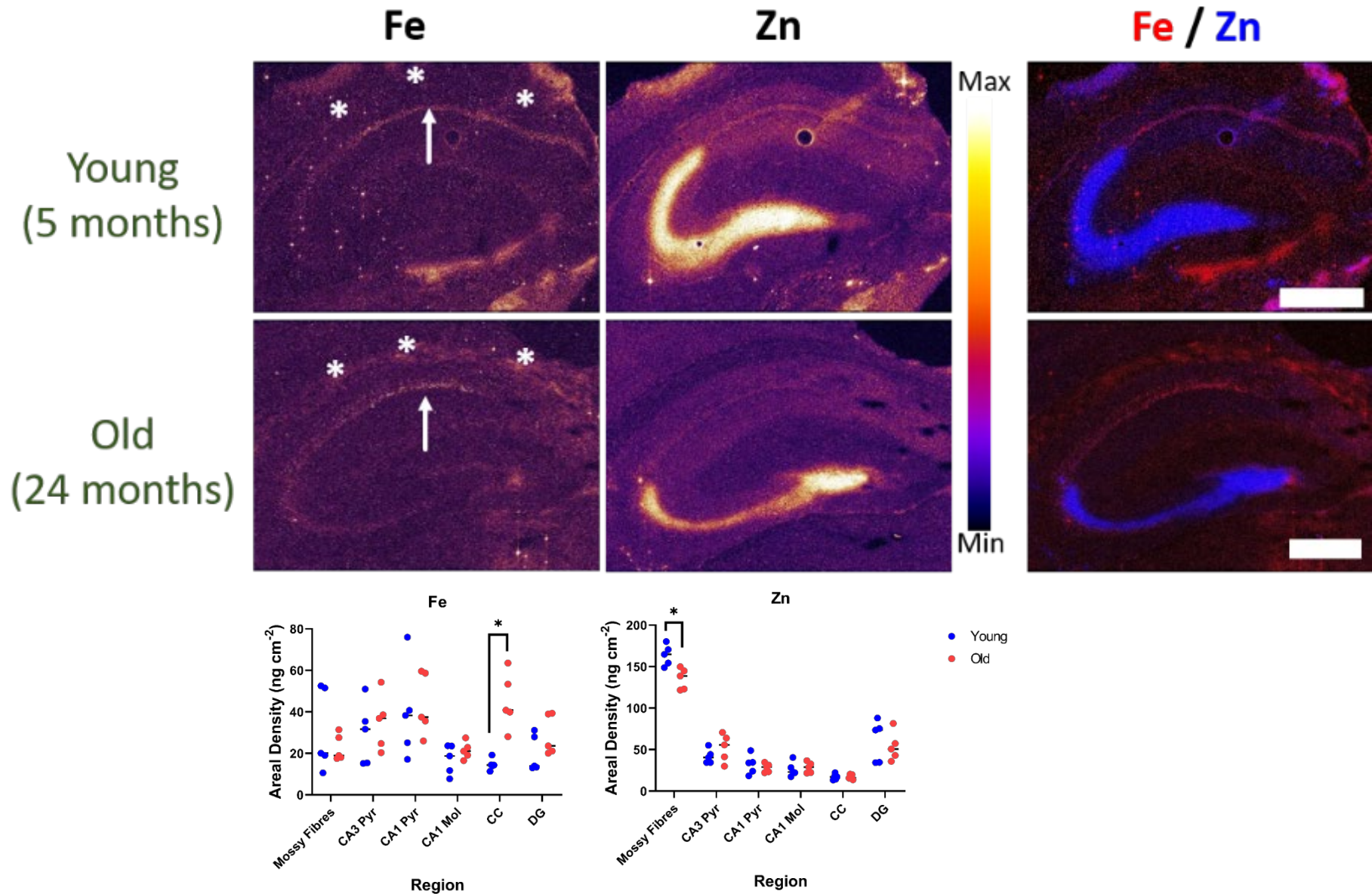
Perls' Stain



Perls' Stain



BRAIN IRON INCREASES IN WHITE MATTER DURING NATURAL AGEING

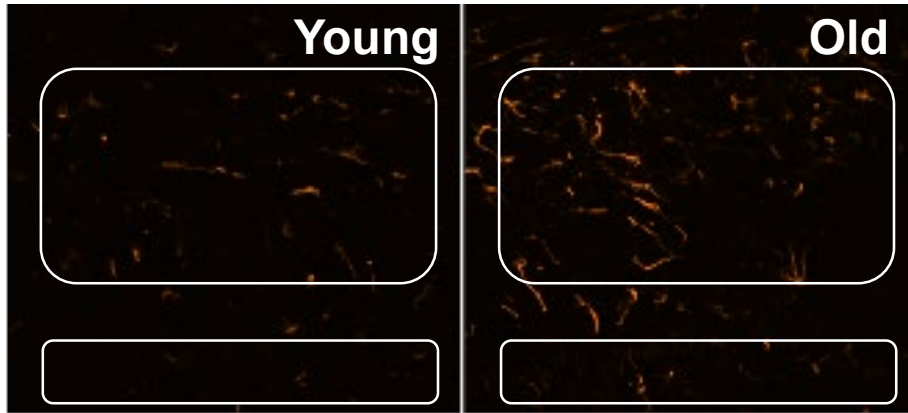


COMBINING XFM WITH IMMUNO-FLUORESCENCE

GFAP (astrocytes)

Young

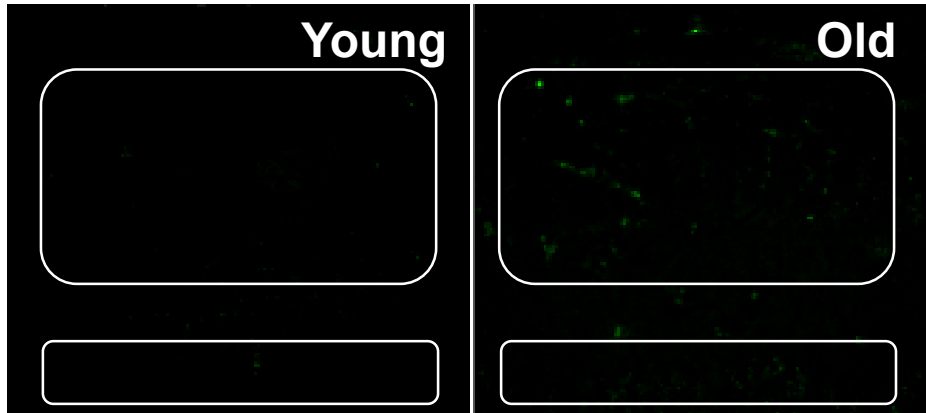
Old



IBA1 (microglia)

Young

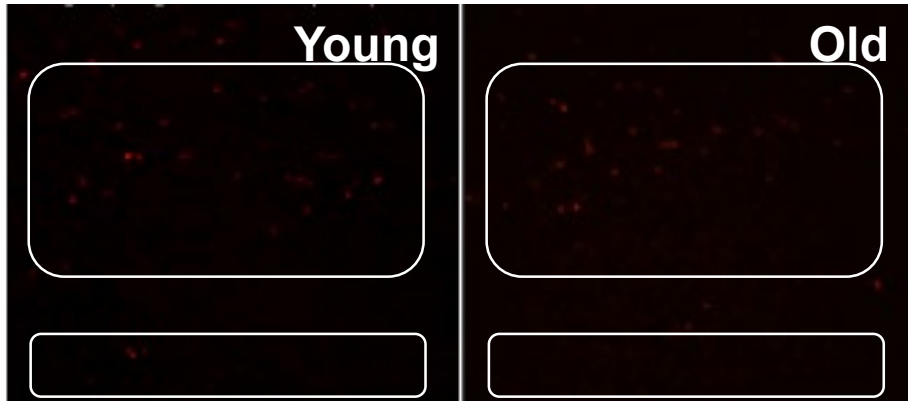
Old



Olig2 (oligodendrocytes)

Young

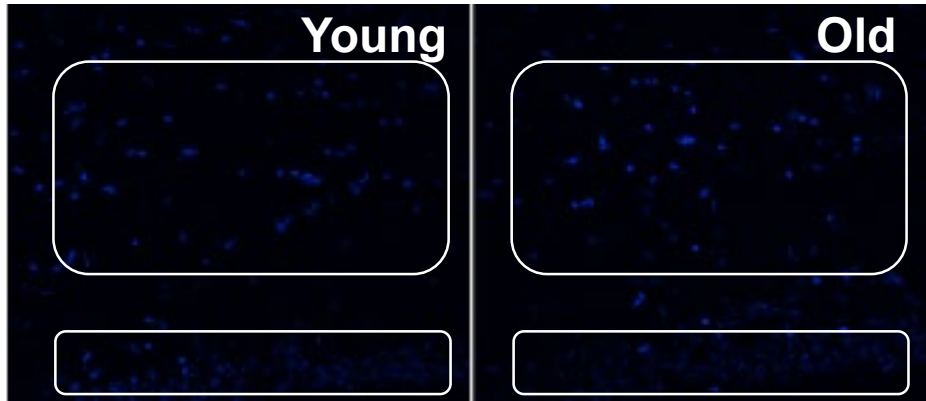
Old



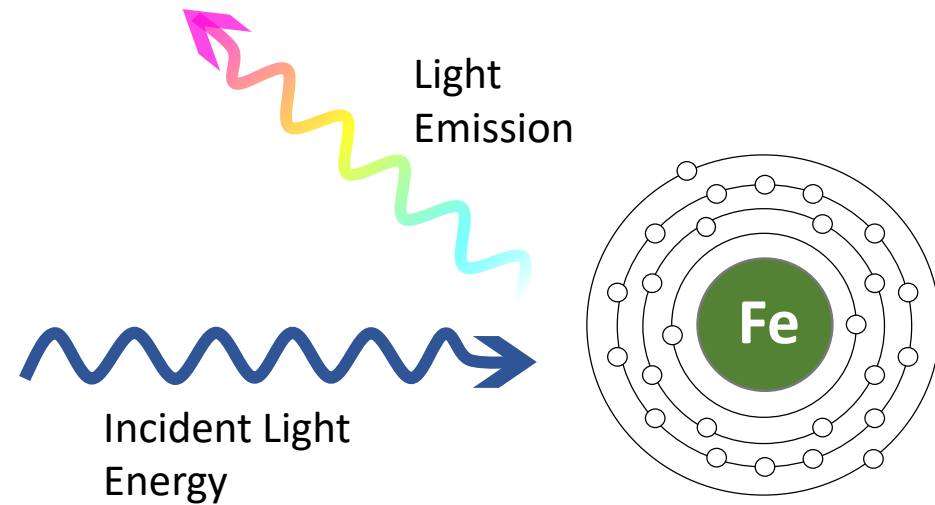
DAPI (nuclei)

Young

Old

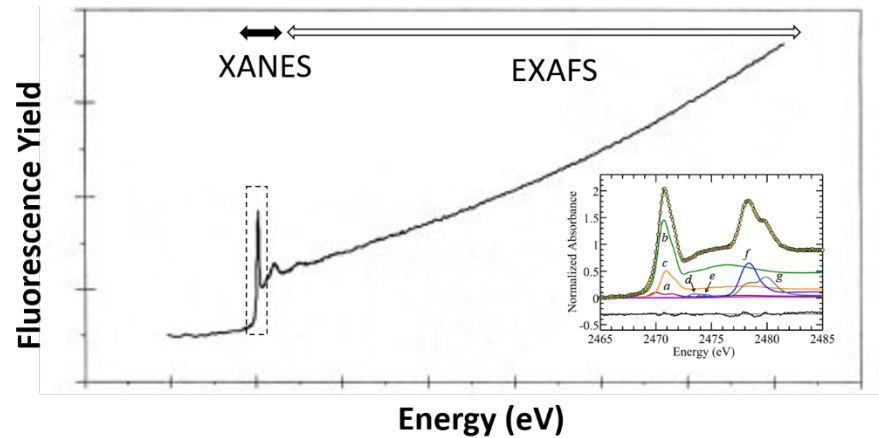


CHEMICALLY SPECIFIC XANES-IMAGING



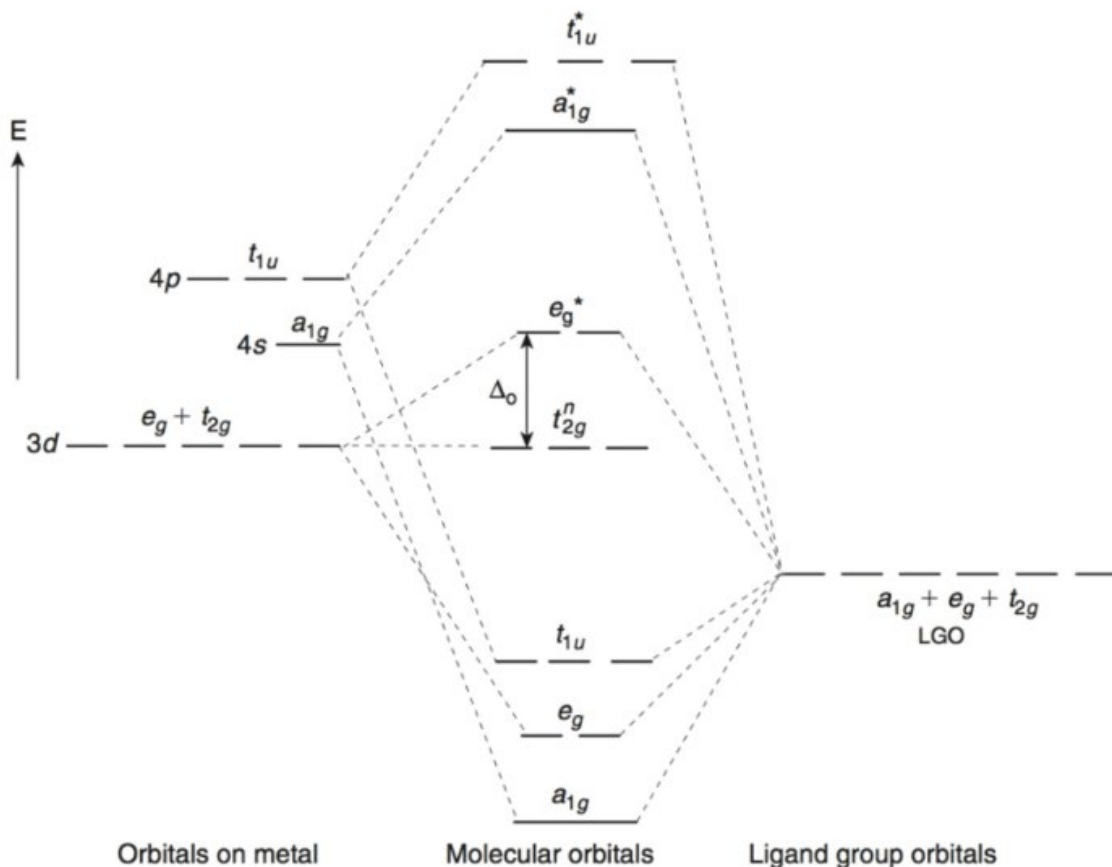
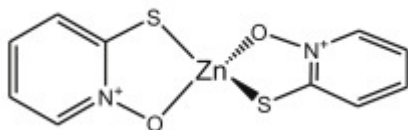
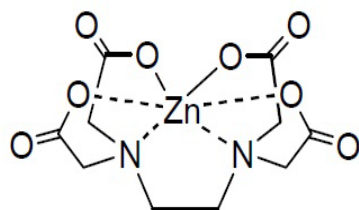
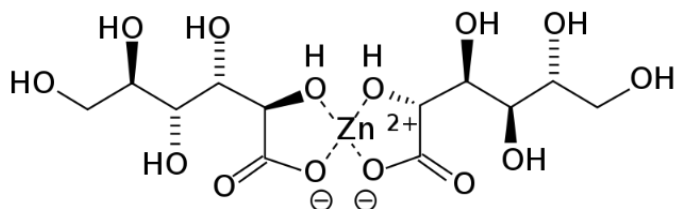
EXAFS: Theory is well understood, **Slow!**

XANES: Theory is not as well understood **Fast!**



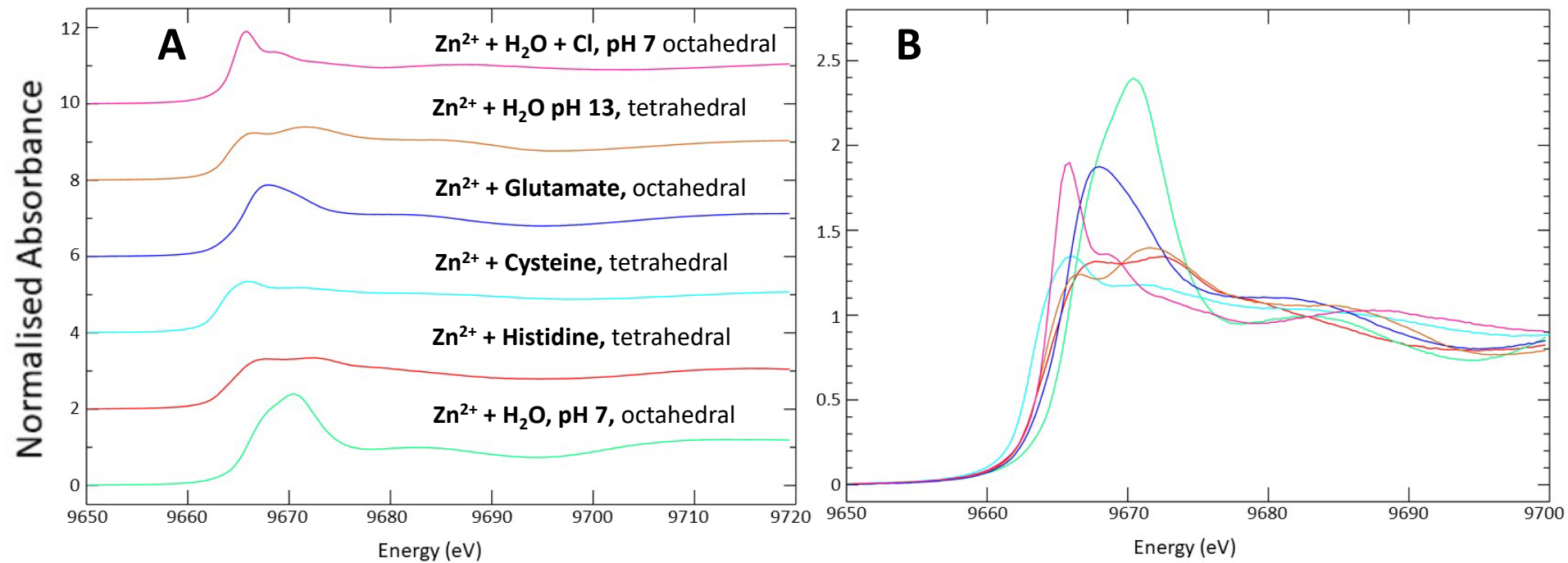
DEVELOPMING ZINC XANES IMAGING

XANES spectroscopy should be highly sensitive to the coordination geometry, number of ligands, and ligand type

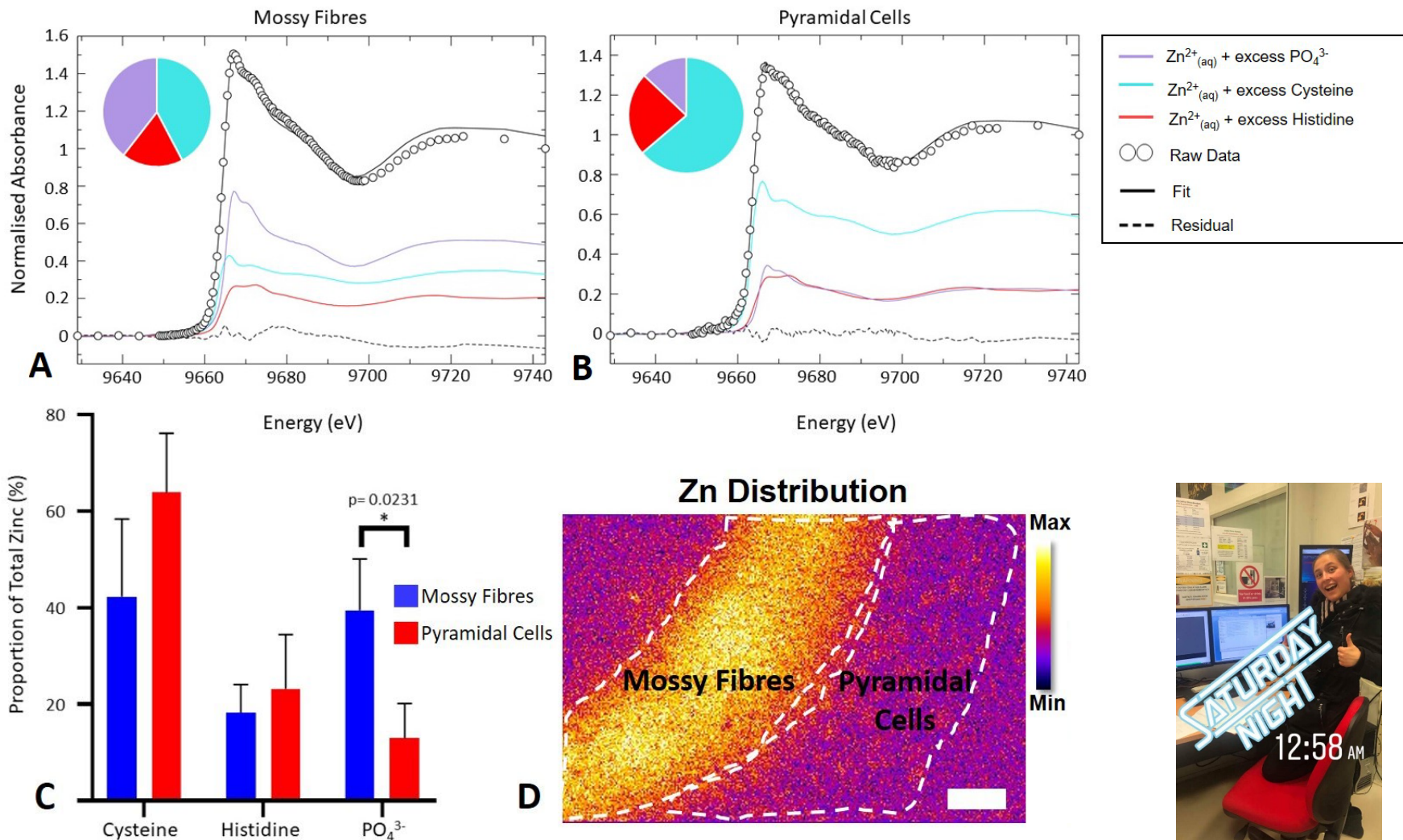


IS THE ZINC K-EDGE SENSITIVE TO COORDINATION CHEMISTRY? (YES)

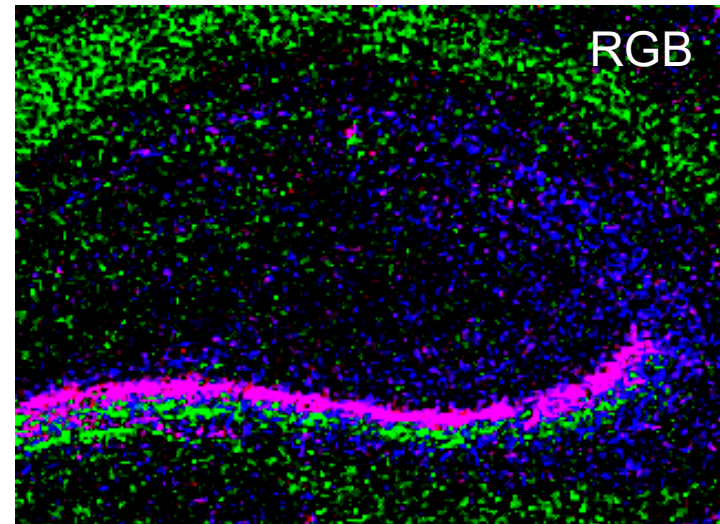
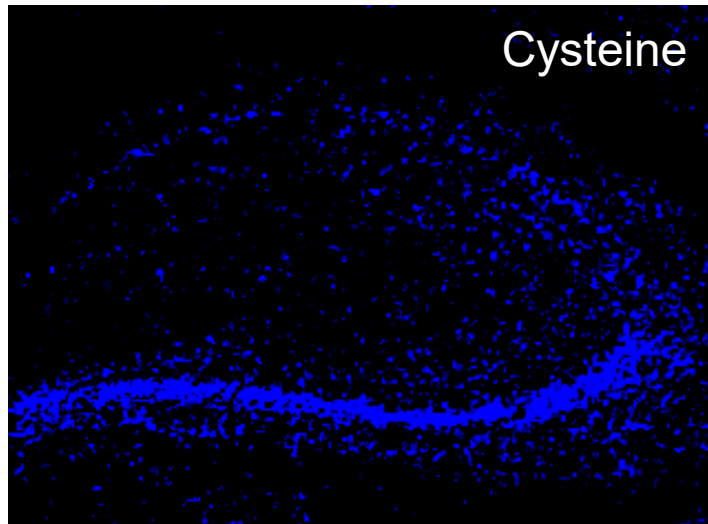
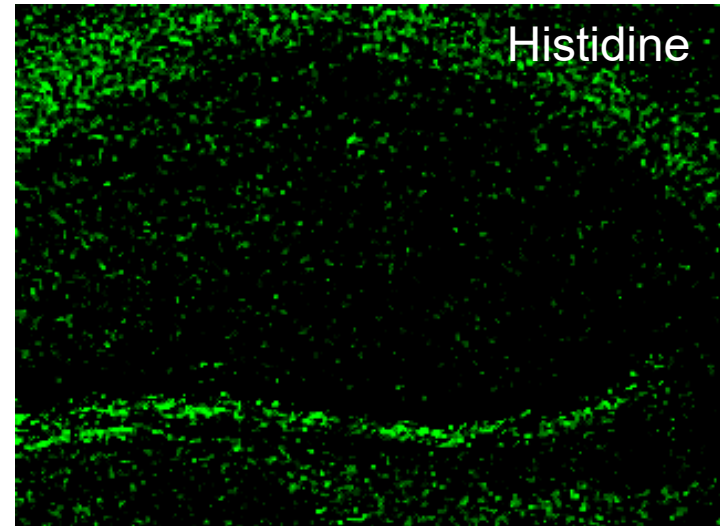
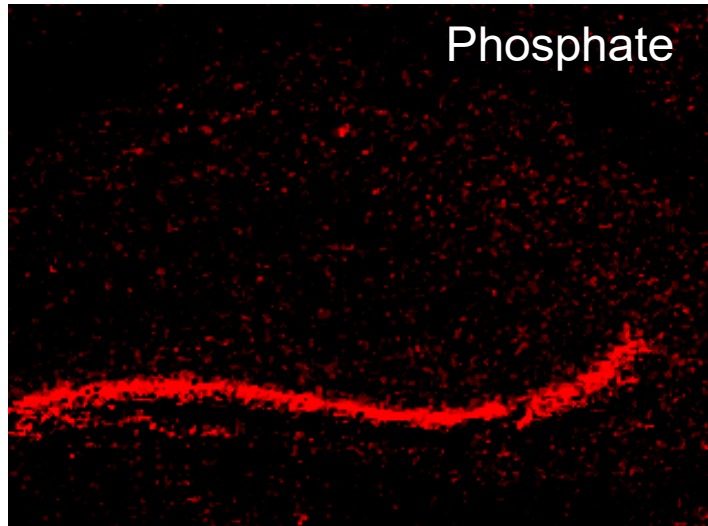
Modelling Zn Bonding in Biological Environments: Zn^{2+} in presence of excess biological ligands



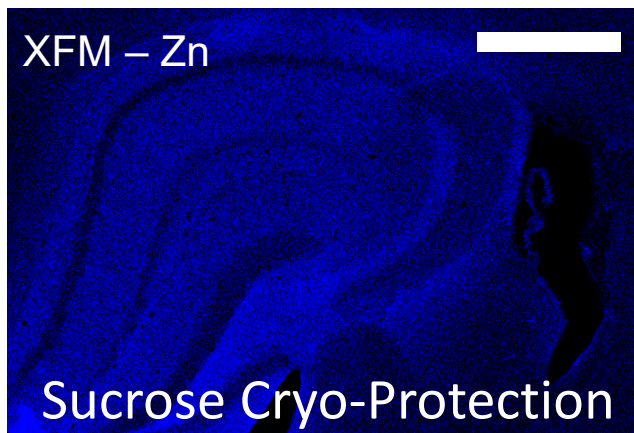
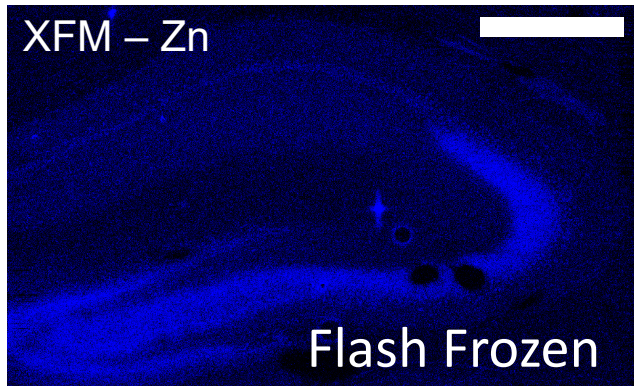
ZINC SPECIATION IN BRAIN TISSUE



CHEMICALLY SPECIFIC XANES ZINC IMAGING



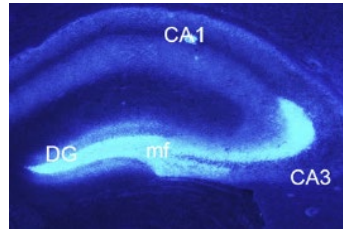
A CAUTIONARY TALE: SAMPLE PREPARATION & ANALYSIS CONDITIONS ARE CRITICAL!!



Zn

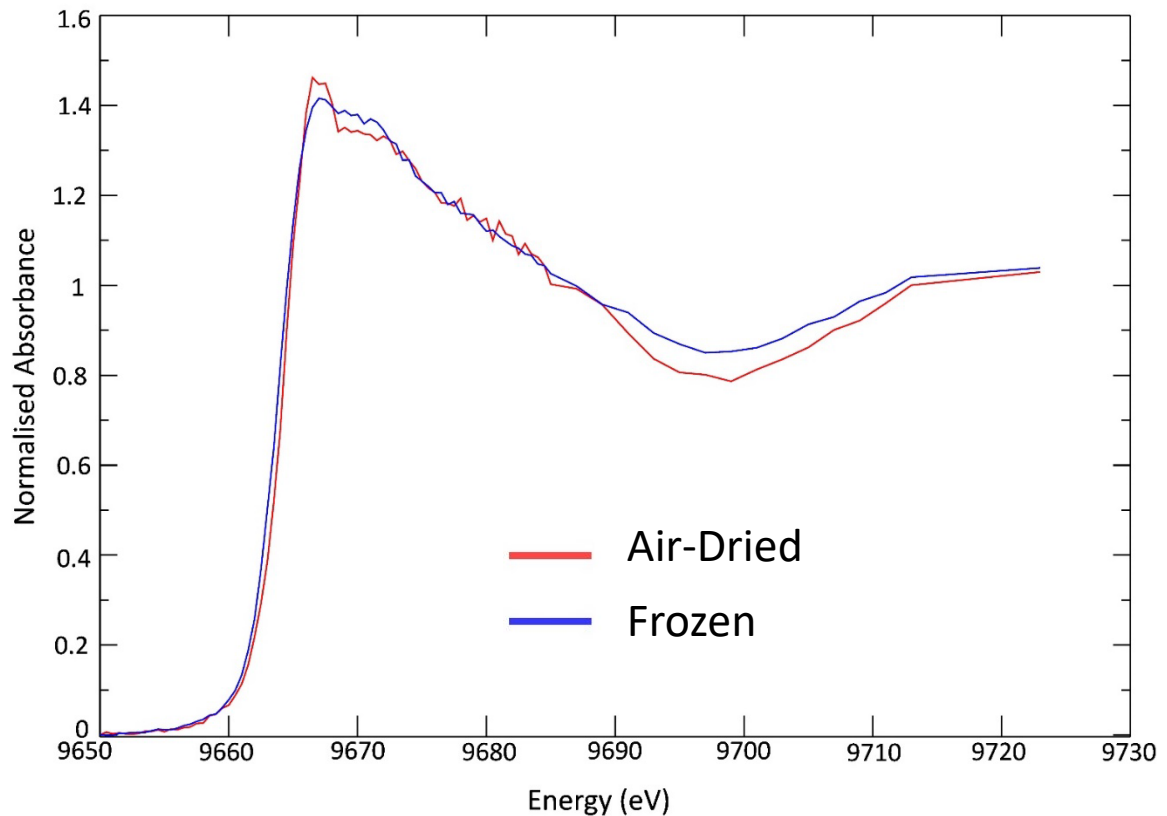
Max Min

Scale = 500 μ m



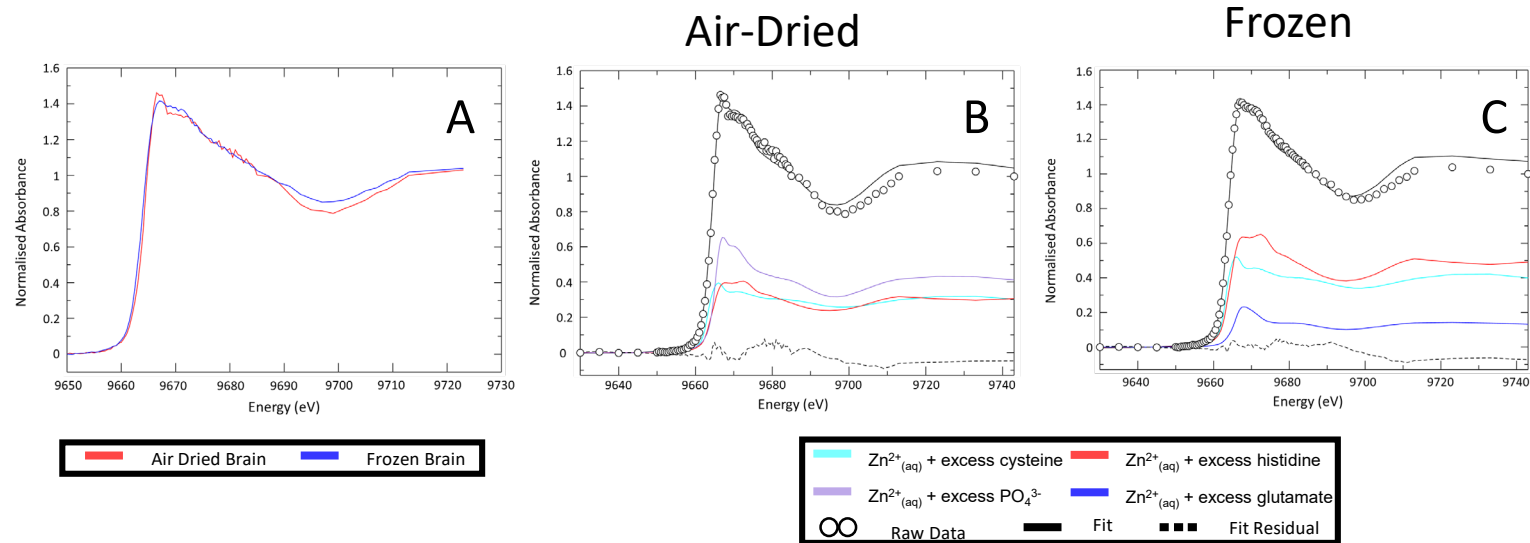
Lee, Joo-Yong, et. al., *Brain research* 1418, 12-22, **2011**

Hollings et. al., *JAAS*, 35, 2498-2508. **2020**

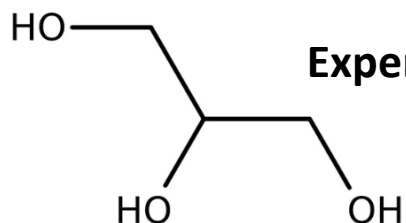


EFFECT OF AIR-DRYING

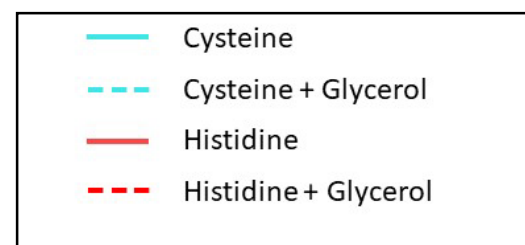
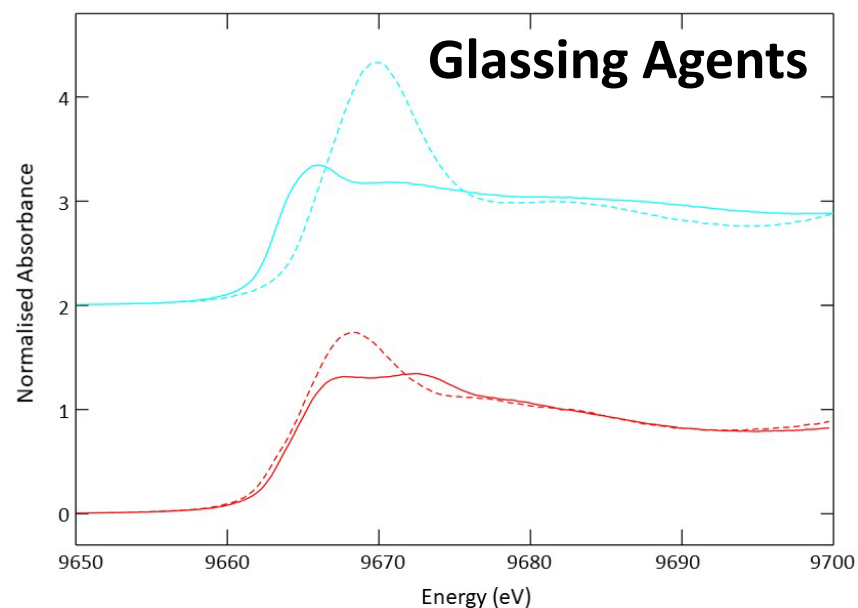
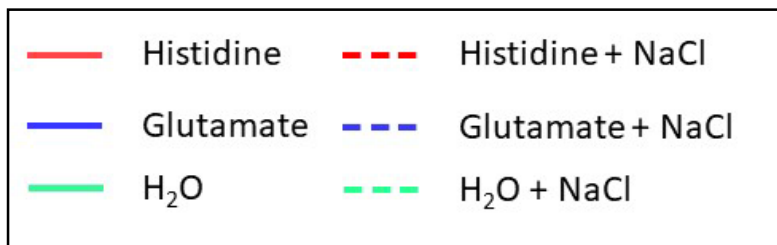
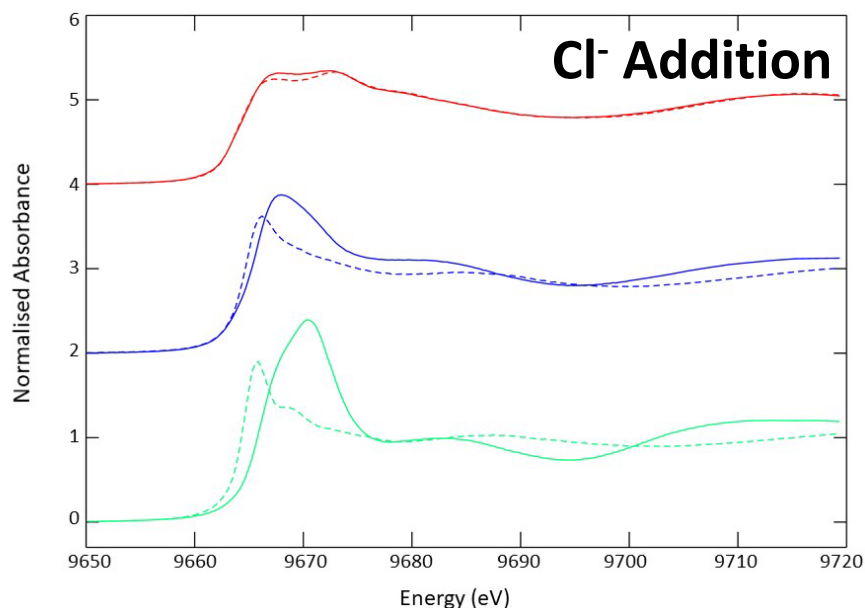
Air-drying tissue sections changes Zn^{2+} speciation, favouring coordination of Zn^{2+} with phosphates (not observed in frozen tissues)



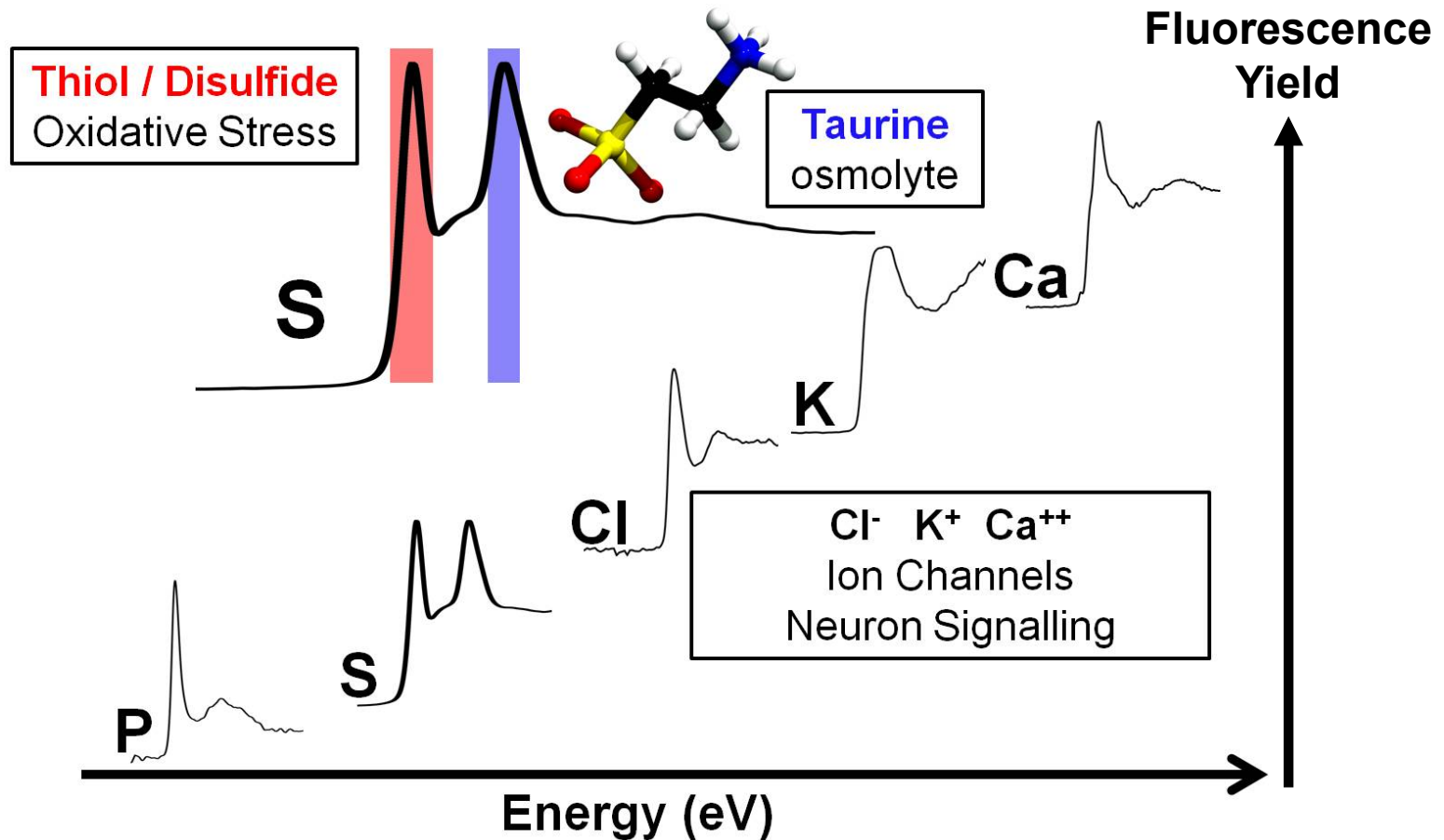
A CAUTIONARY TALE: SAMPLE PREPARATION & ANALYSIS CONDITIONS ARE CRITICAL!!



**Experimental considerations for the XANES library:
buffer ions, and “glassing agents”**

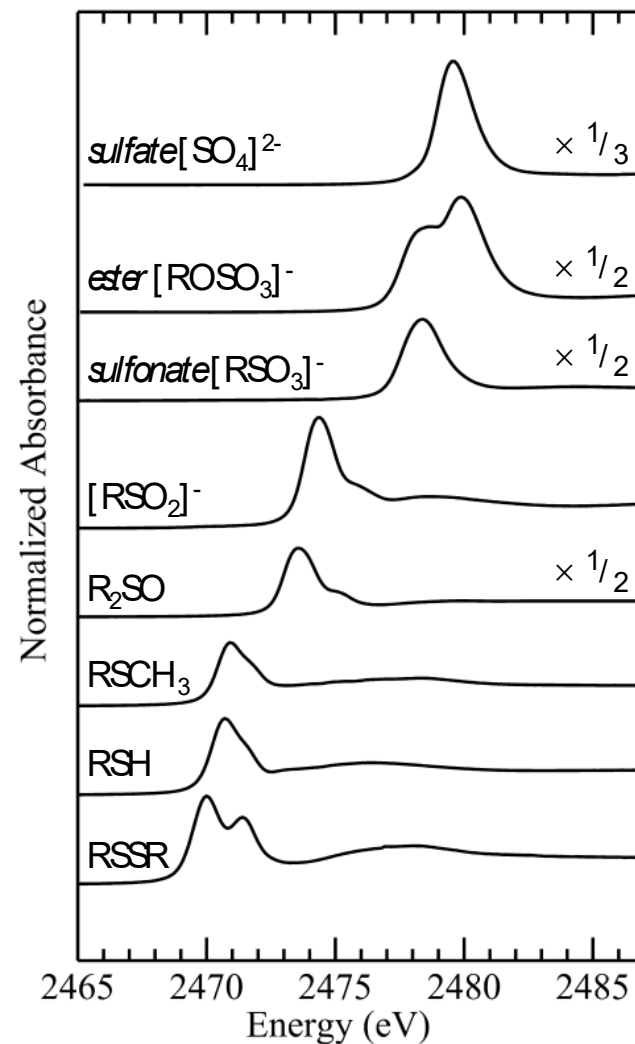
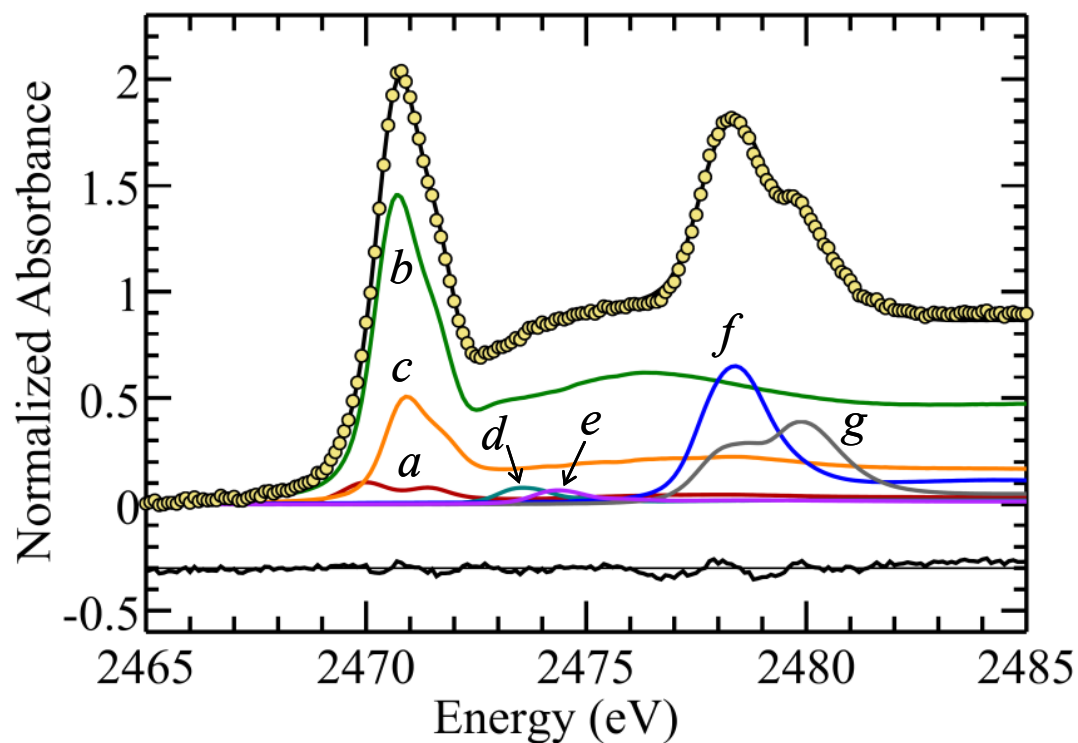


MEDIUM ENERGY X-RAY ABSORPTION SPECTROSCOPY (MEX) @ THE SYNCHROTRON

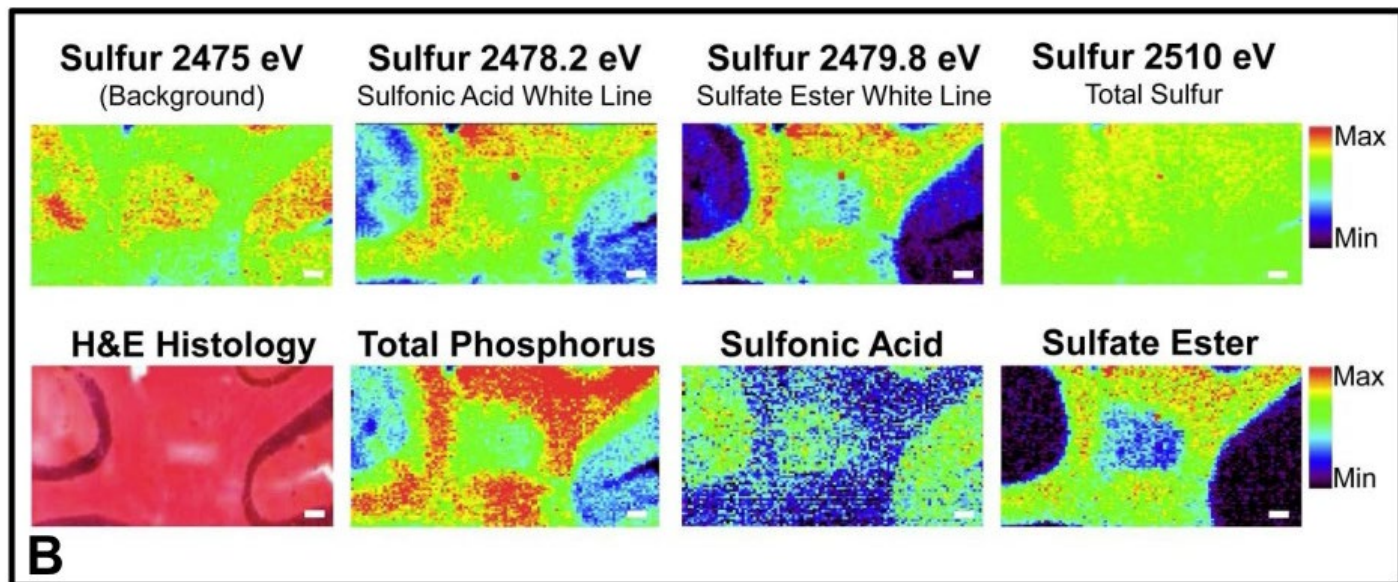
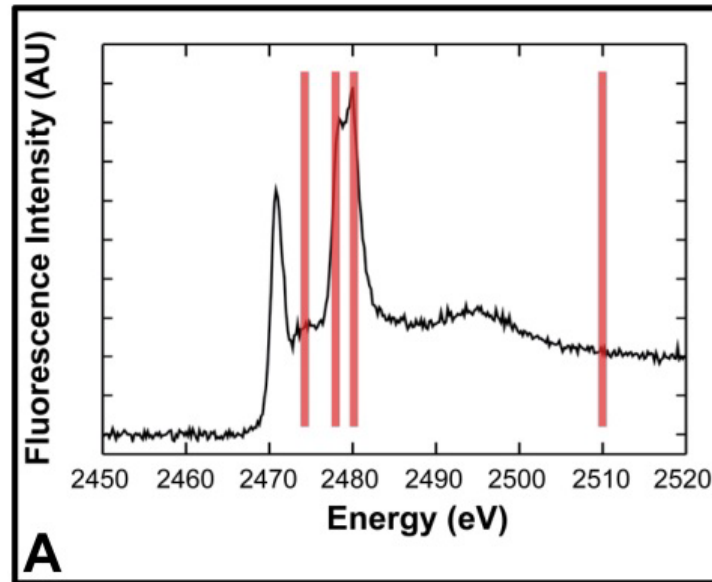


MEDIUM ENERGY X-RAY ABSORPTION SPECTROSCOPY

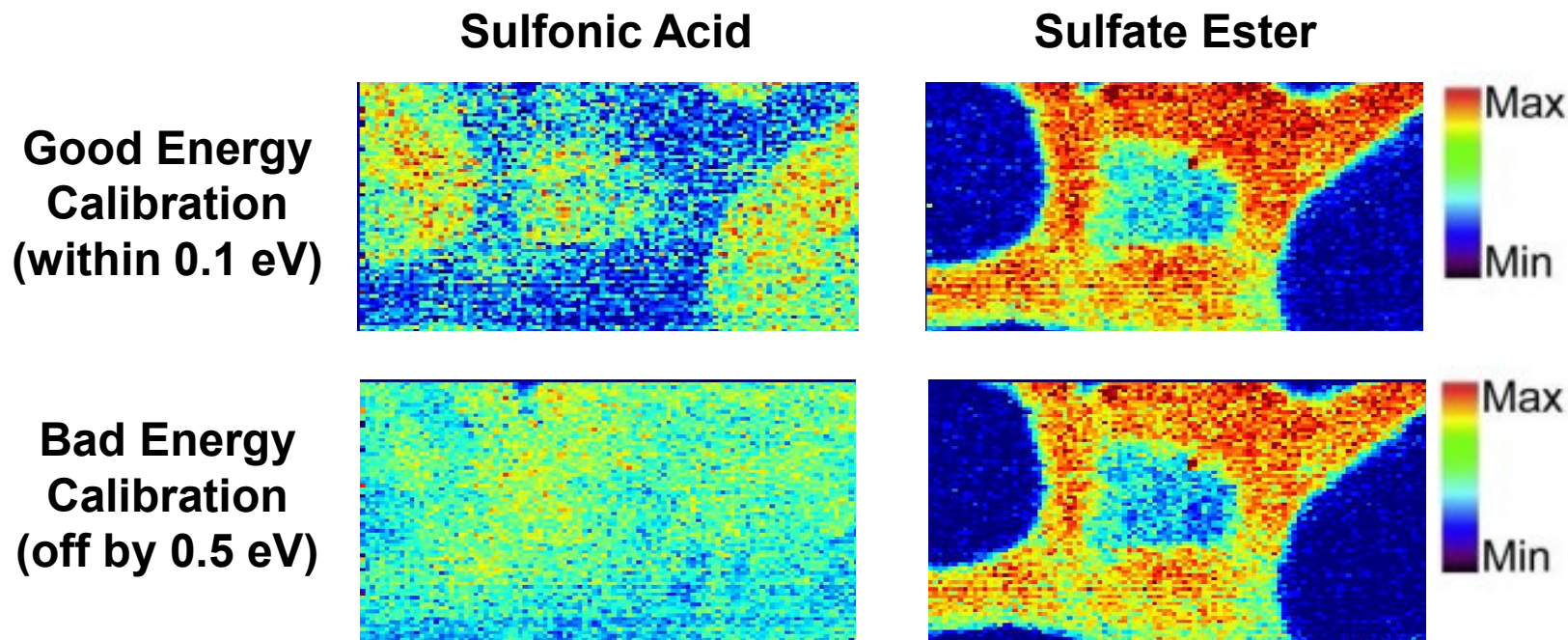
MEX Beamline: BRIGHT



MEDIUM ENERGY X-RAY ABSORPTION SPECTROSCOPY



Pit falls with chemically specific XFM



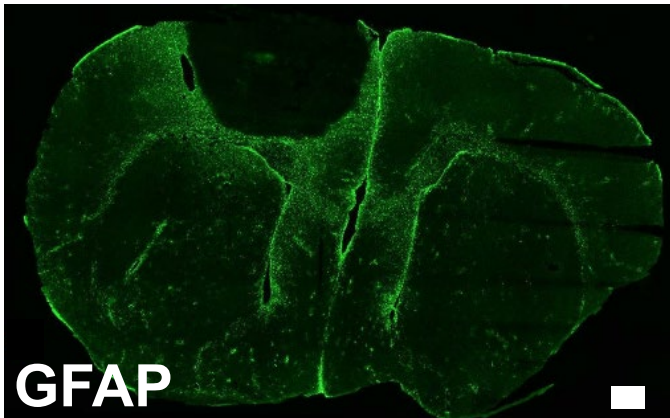
Energy Calibration & Stability is Critical!

MEX: IS THERE AN ISCHEMIC PENUMBRA IN THE MURINE PHOTOTHROMBOTIC STROKE MODEL?

H&E



GFAP

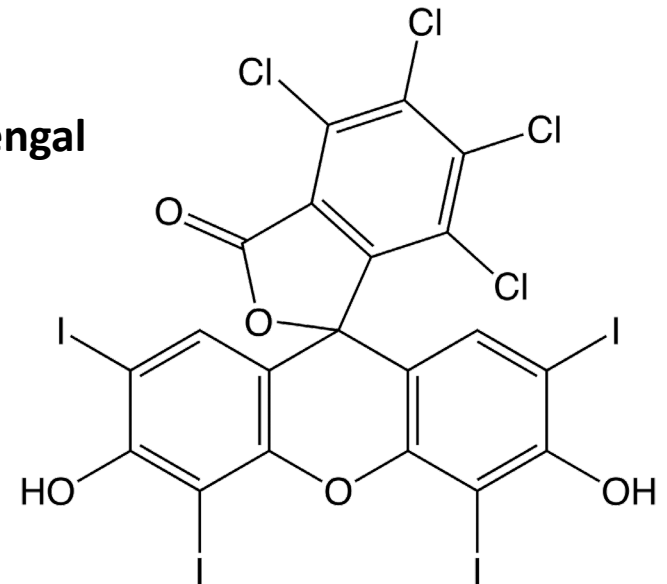


Scale = 500 μ m



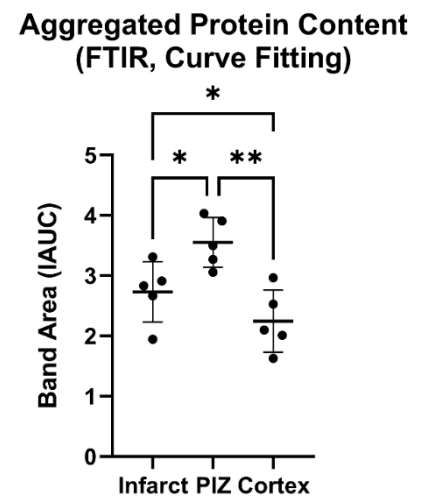
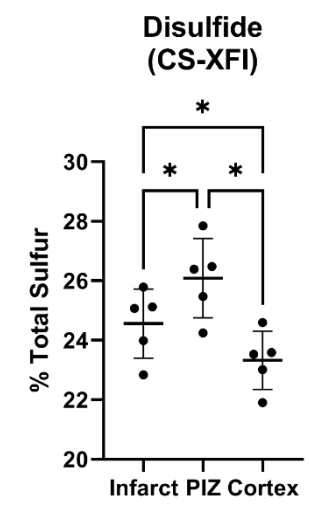
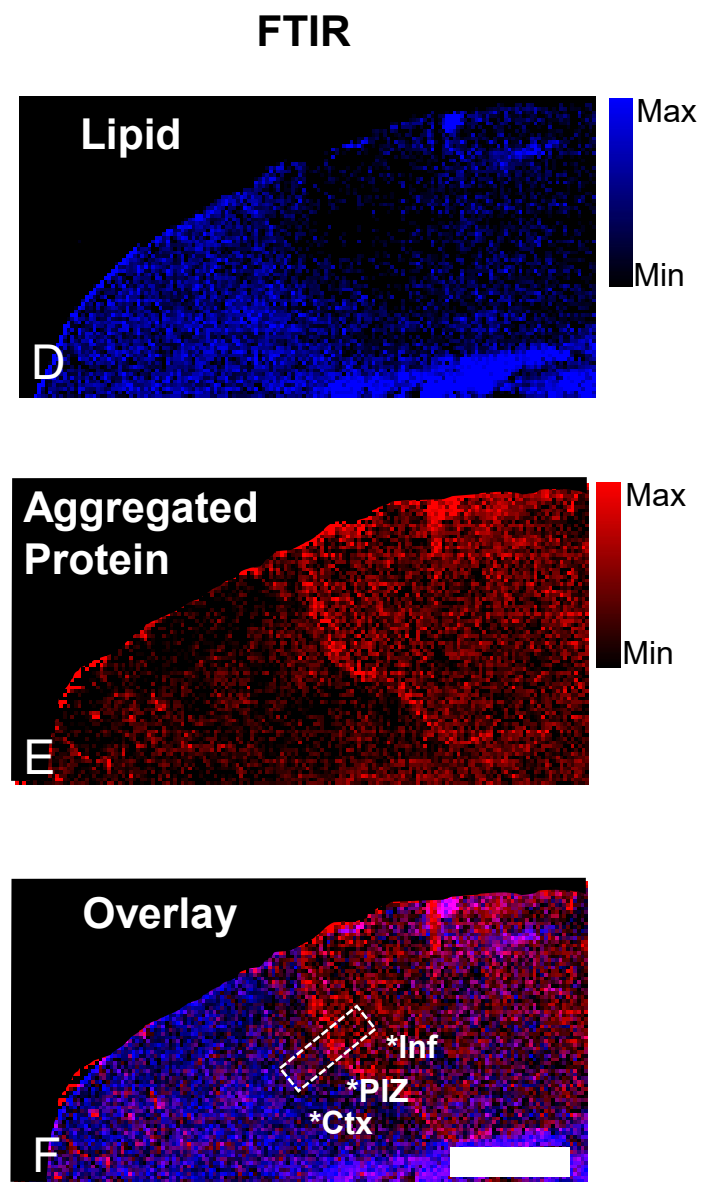
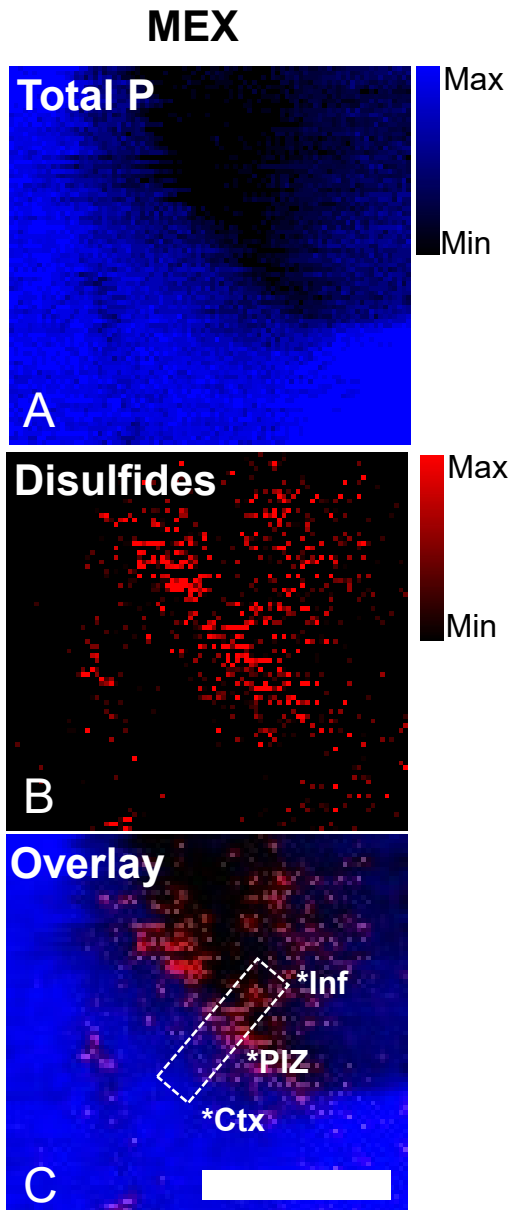
Saskatchewan Cerebrovascular Unit

Rose Bengal

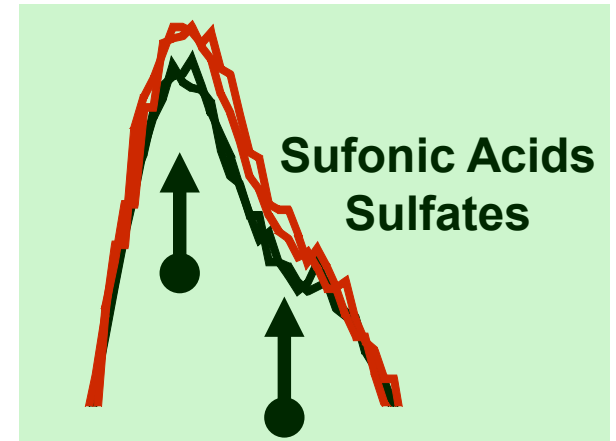
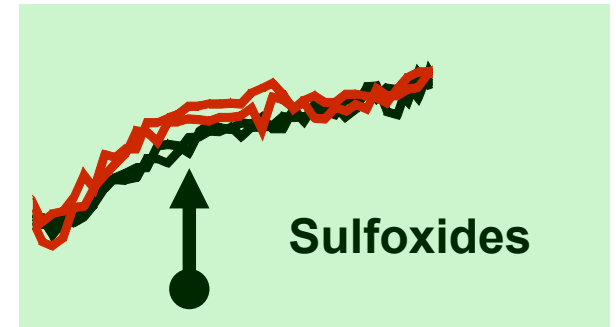
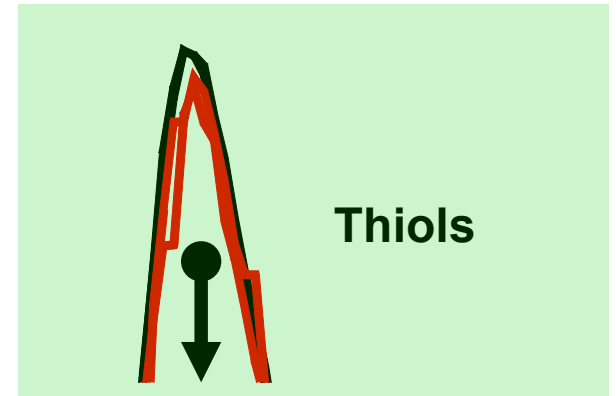
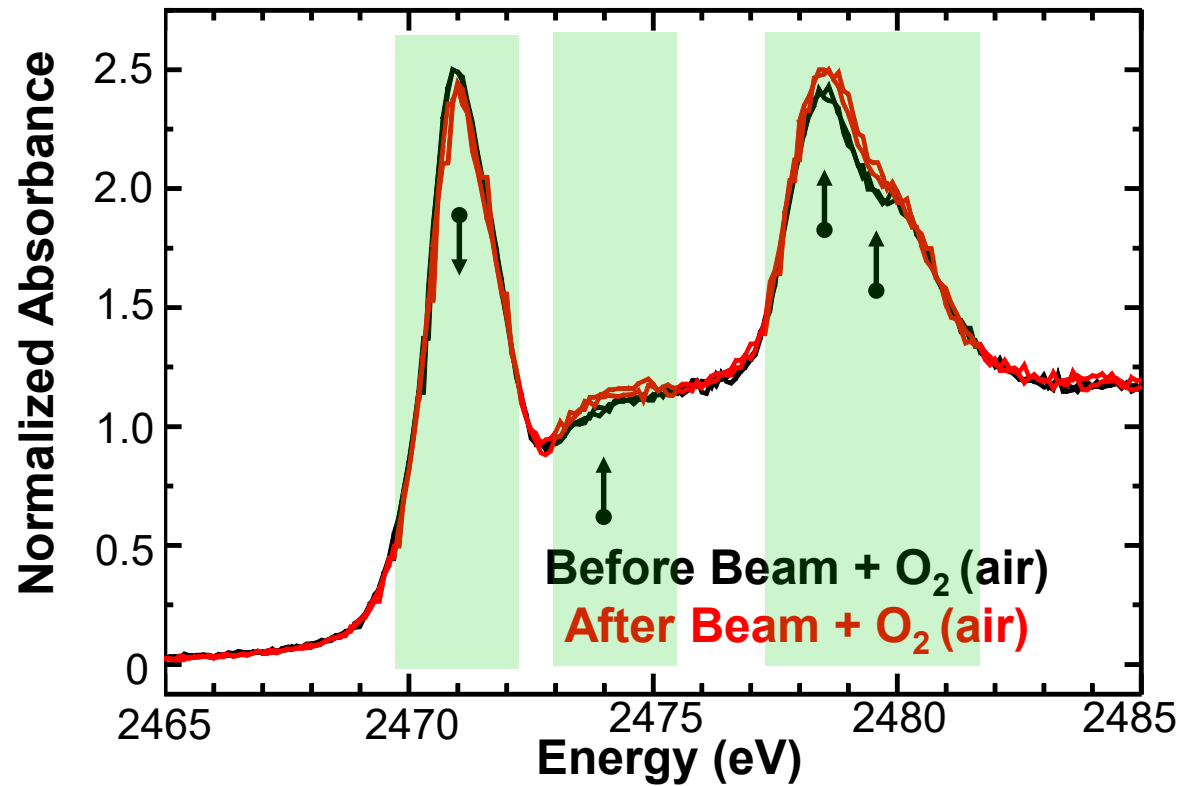


Is there an ischemic penumbra in the photothrombotic stroke model?

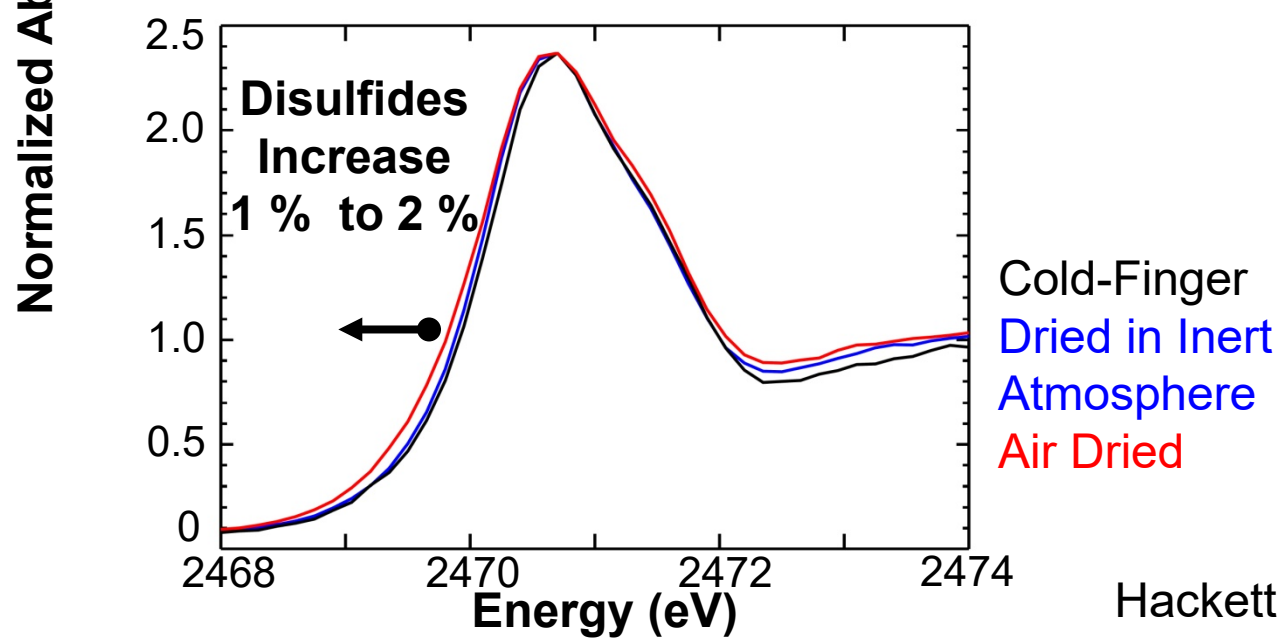
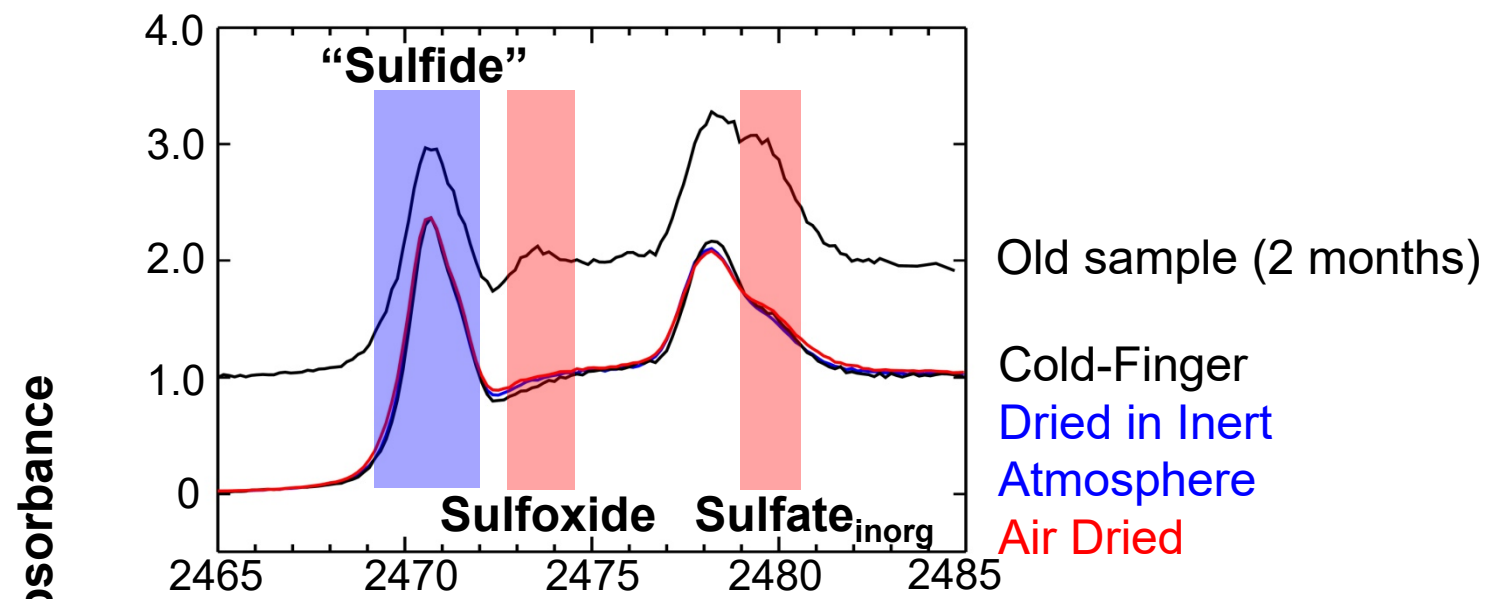
MEX: IS THERE AN ISCHEMIC PENUMBRA IN THE MURINE PHOTOTHROMBOTIC STROKE MODEL?



X-RAY PHOTO-DAMAGE

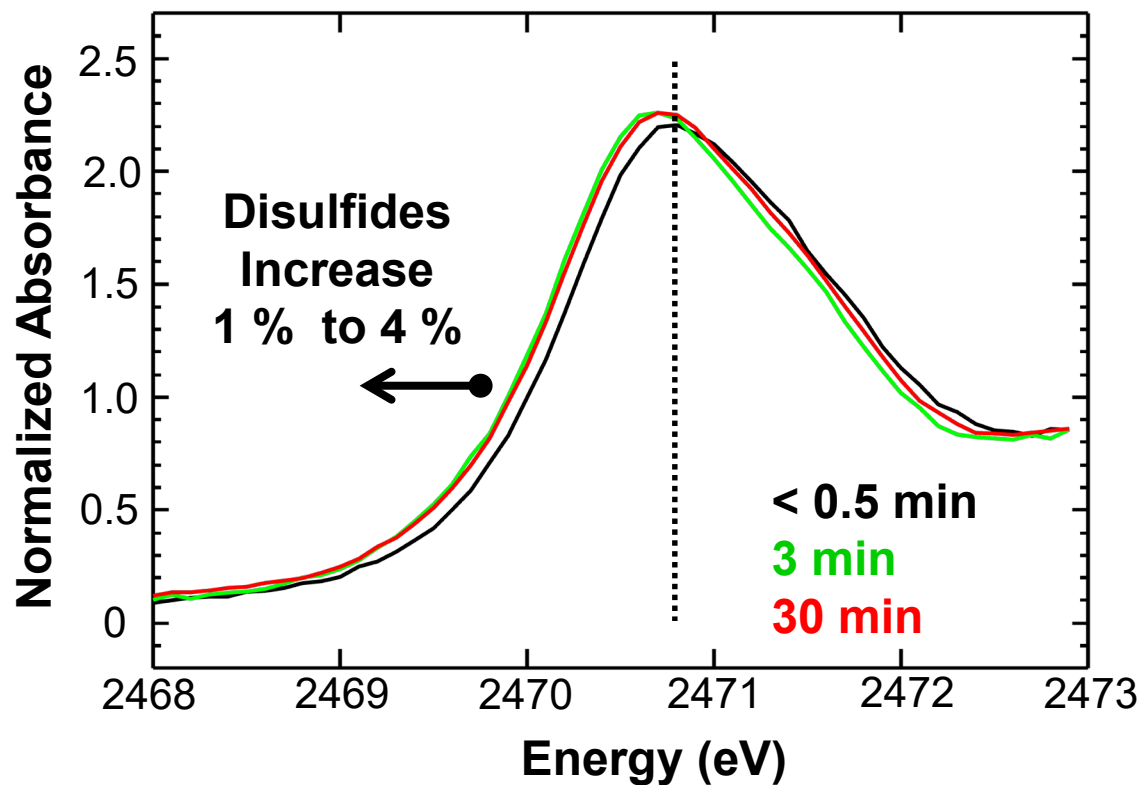


Effects of sample preparation



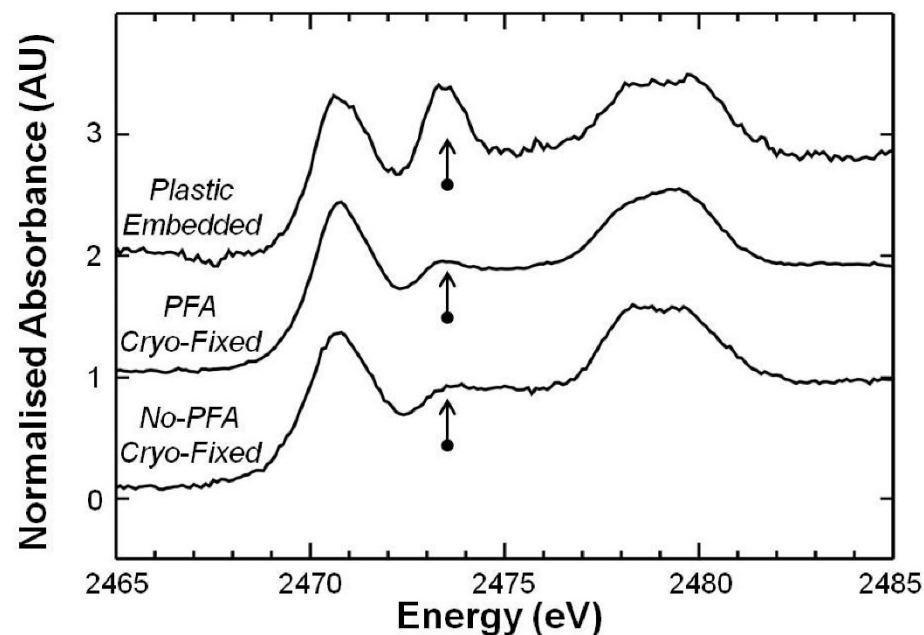
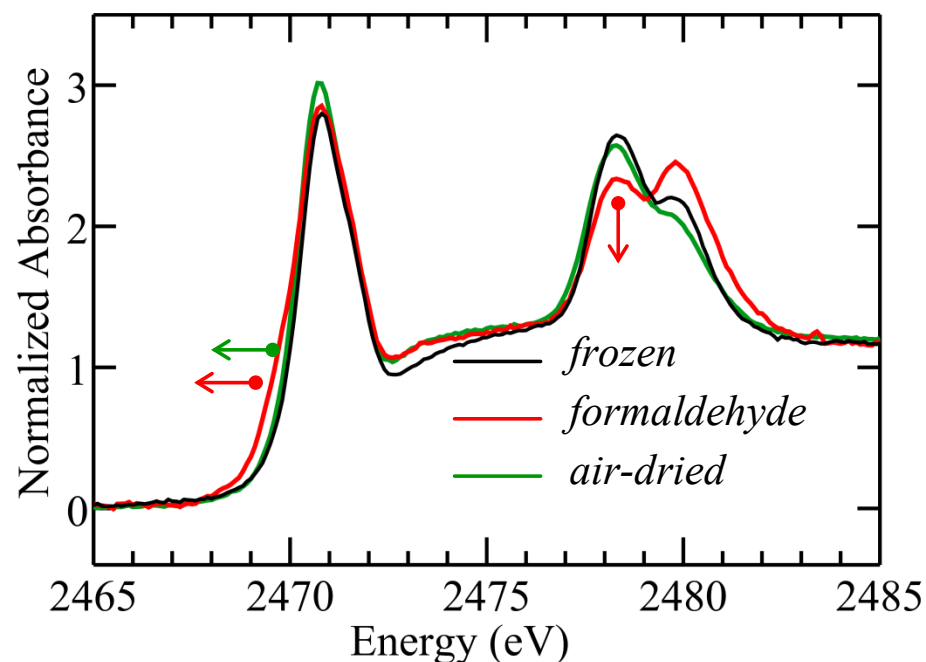
EFFECTS OF POST-MORTEM INTERVAL

The time it takes to sacrifice an animal and remove its brain (30 seconds – 3 minutes) induces ischemia and oxidises thiols to disulfides



SAMPLE PREPARATION – DON'T USE CHEMICAL FIXATION

Chemical fixation = sulfonic acid leaching & thiol oxidation
Paraffin or plastic embedding = massive sulfur oxidation



Hackett et al. (2012) ACS Chemical Neuroscience

Hackett et al. (2016) ACS Biochemistry

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CURTIN HEALTH
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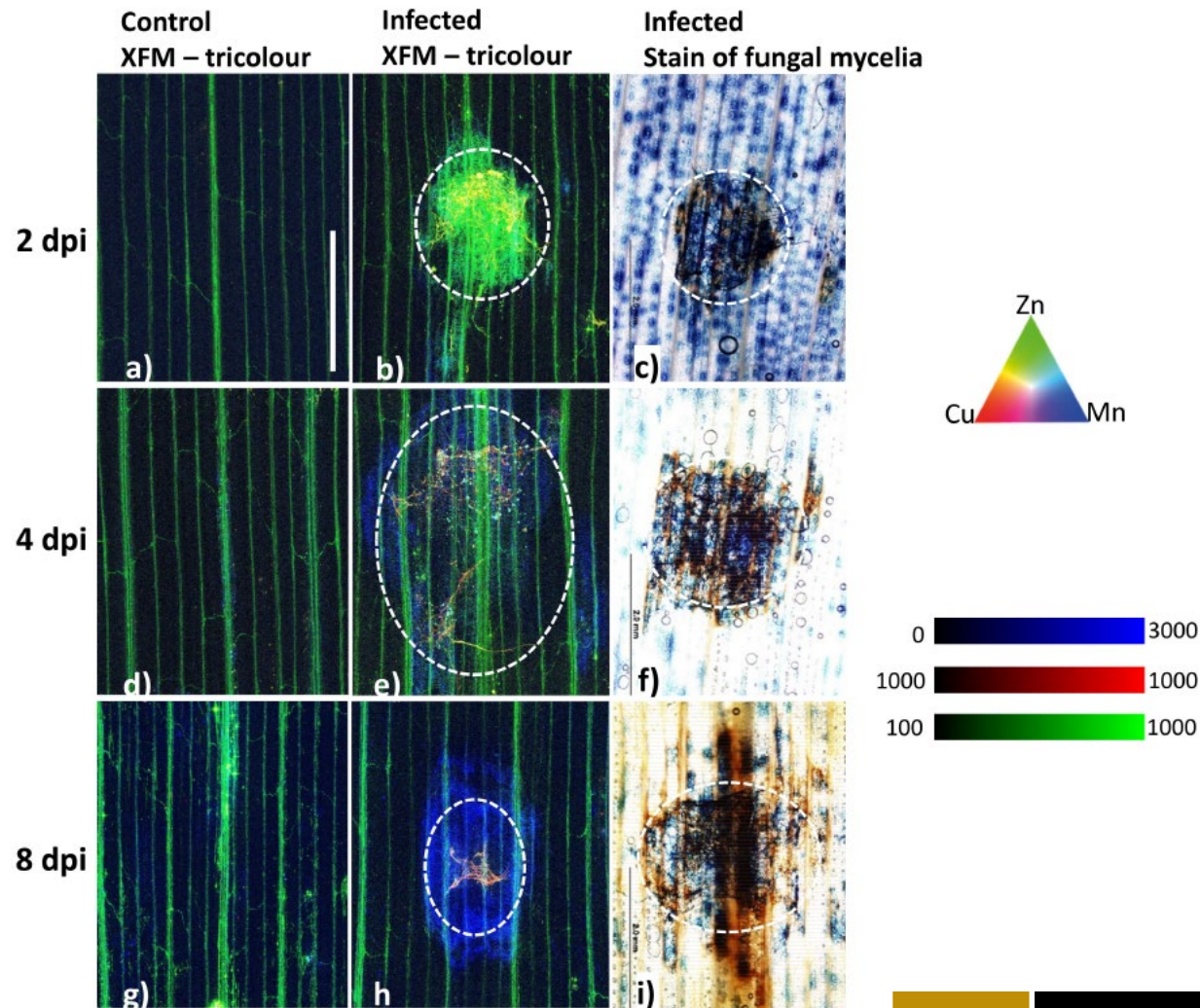


Australian Government

Australian Research Council



4. THE ROLE OF METALS IN WHEAT FUNGAL DISEASE



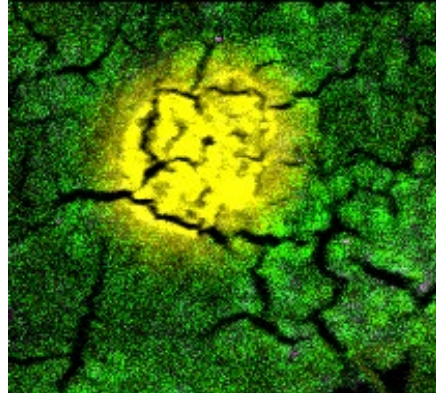
Slide courtesy of Fatima Naim and Mark Gibberd



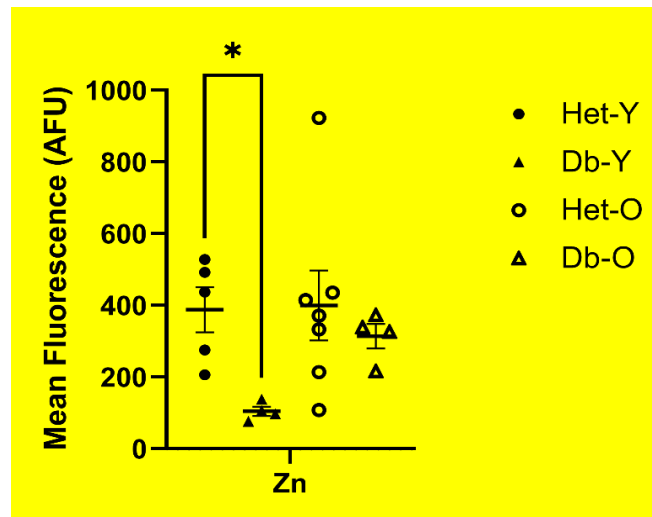
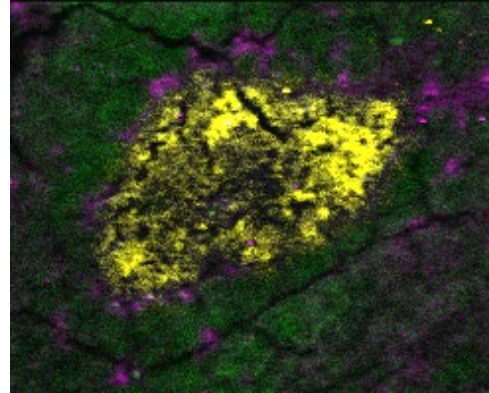
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4. ZINC HOMEOSTASIS IN THE PANCREAS

Young (WT)

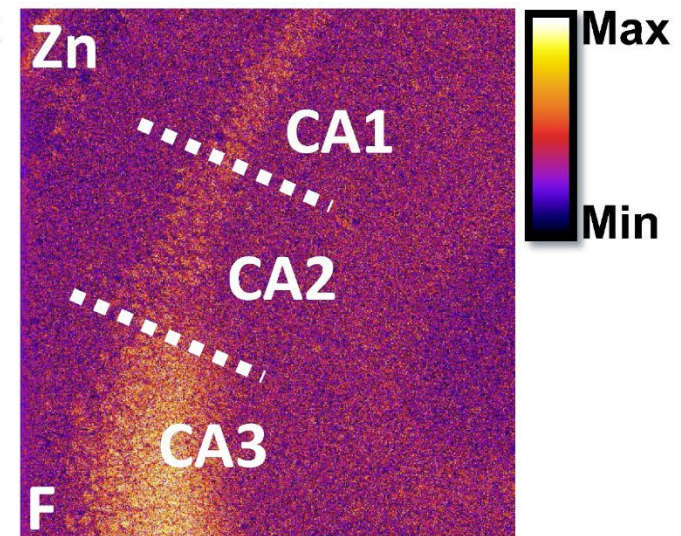
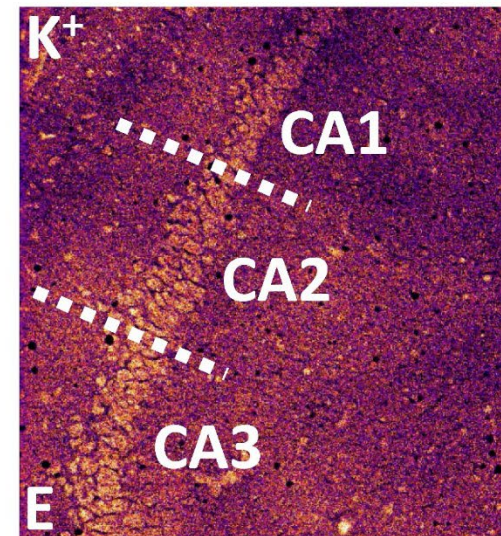
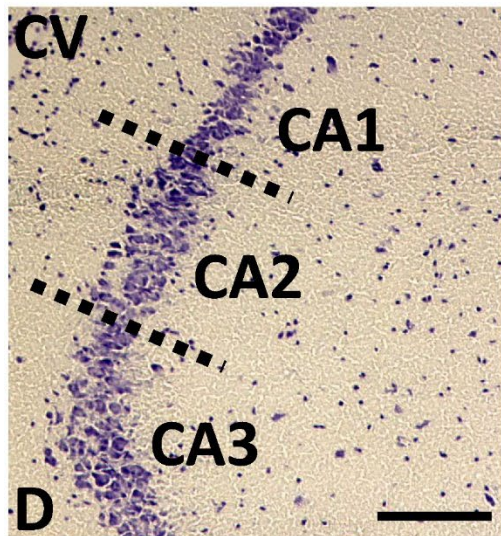
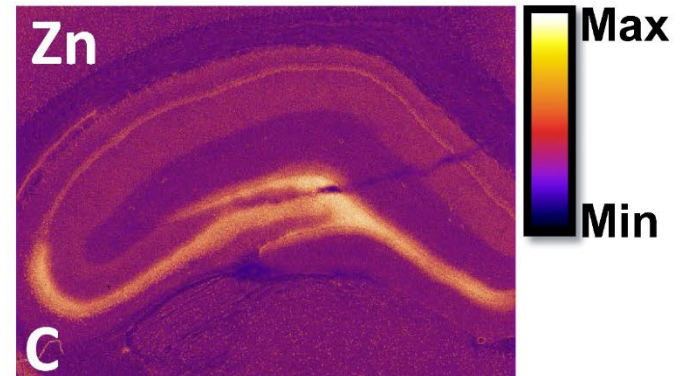
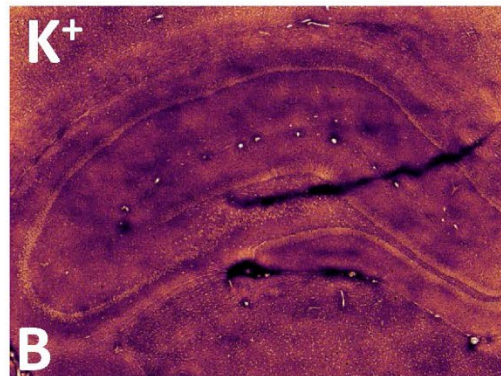
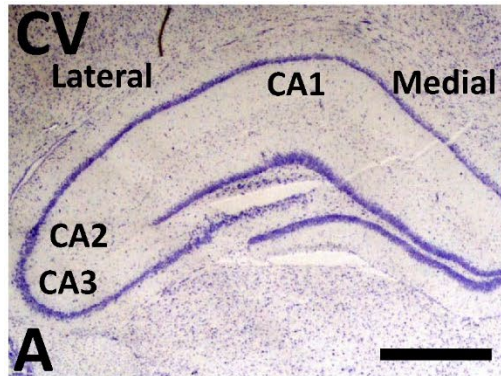


Young (dbdb)

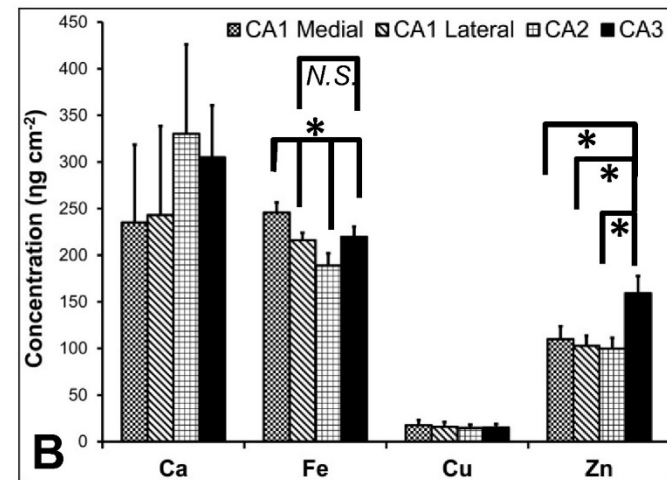
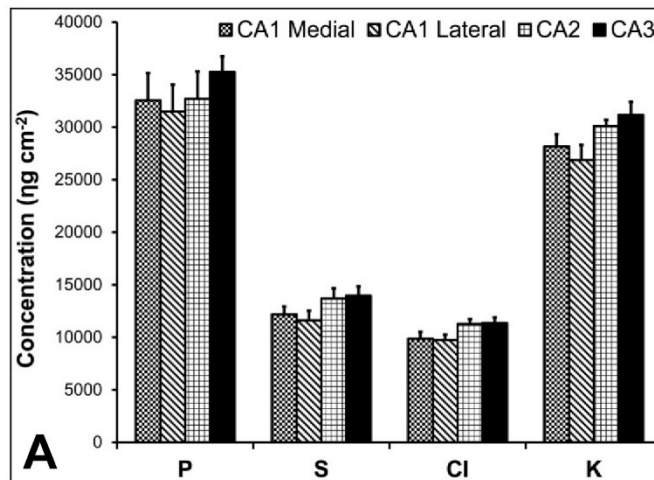
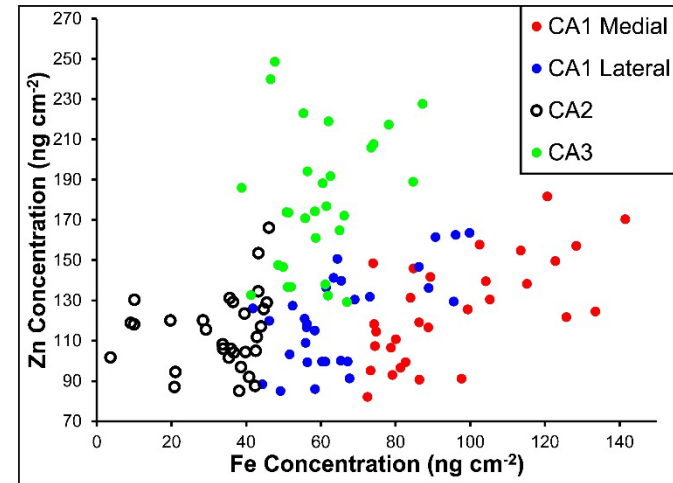
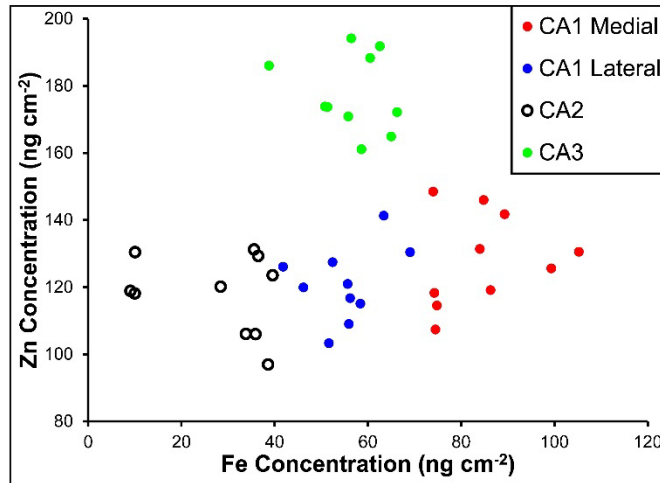


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ANATOMICAL DEMARCATION FROM ELEMENTAL MAPS



2. DETERMINING ELEMENTAL PROFILES IN RAT HIPPOCAMPAL PYRAMIDAL NEURONS



4. ELEMENTAL MAPPING LATENT FINGERMARKS



X-ray Fluorescence Microscopy

Aim: Image the spatial distribution of elemental components within a natural fingerprint deposit

Approach: 8 donors deposit natural fingerprints on silicon nitride slides or Mylar

Elements detected at trace levels:

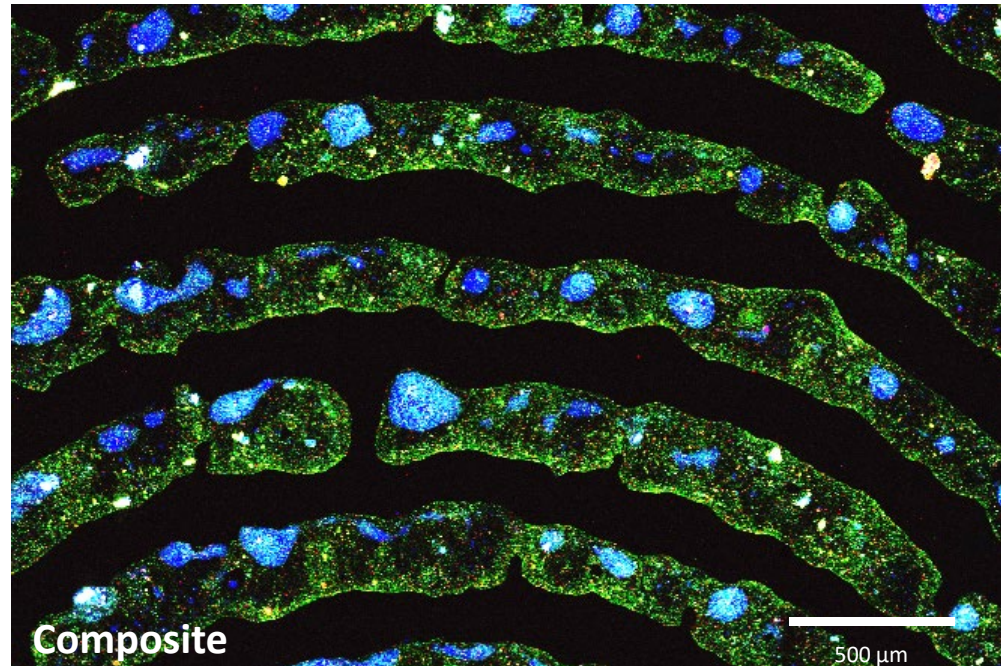
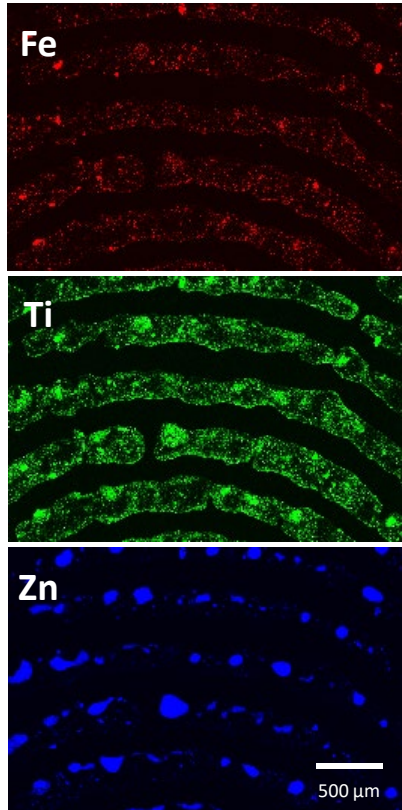
Calcium, Chloride, Copper, Iron, Nickel, Potassium, Titanium, Zinc



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4. Elemental mapping latent fingerprints

Co-localisation of metals from proposed exogenous sources

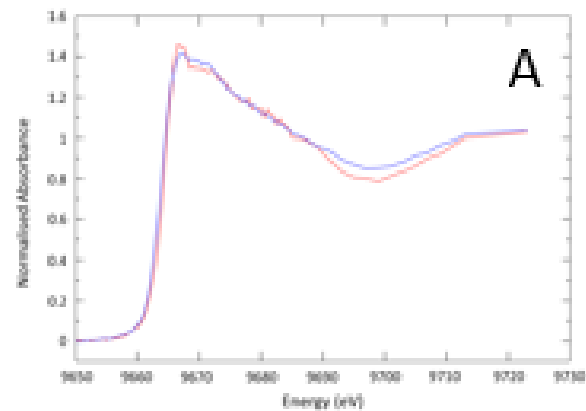


Donor 1 natural fingerprint deposited on silicon nitride

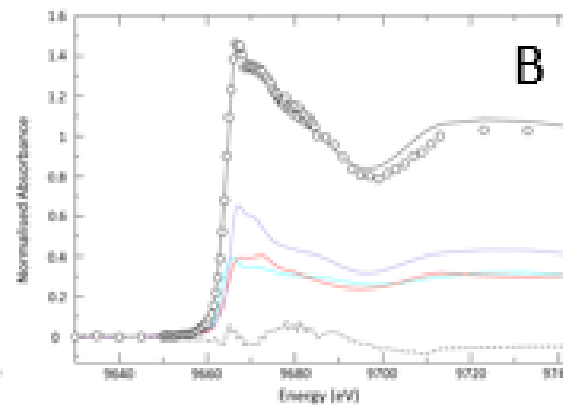


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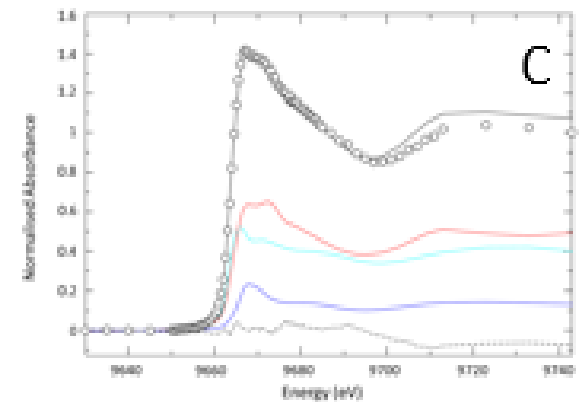
EFFECT OF AIR-DRYING



— Air Dried Brain — Frozen Brain



— $\text{Zn}^{2+}_{(\text{aq})} + \text{excess cysteine}$ — $\text{Zn}^{2+}_{(\text{aq})} + \text{excess histidine}$
— $\text{Zn}^{2+}_{(\text{aq})} + \text{excess PO}_4^{3-}$ — $\text{Zn}^{2+}_{(\text{aq})} + \text{excess glutamate}$
○ Raw Data — Fit ■ Fit Residual



MEDIUM ENERGY X-RAY ABSORPTION SPECTROSCOPY

